Bibliometric Analysis of Published Literature Utilizing Optogenetics as a Technique in Neuroscience: 2010–2020

Dwight Figueiredo^{1,*}, Paulomi Sanghavi²

¹Symbiosis Institute of Health Sciences (SIHS), A Constituent of Symbiosis International (Deemed to be University) (SIU), Pune, Maharashtra, INDIA.

²Department of Biological Sciences (DBS), Tata Institute of Fundamental Research, Mumbai, Maharashtra, INDIA.

ABSTRACT

The optogenetics technique where in light is used to excite or inhibit neurons has uncovered many new brain neuronal circuits causal in or associated with routine day-to-day physiological functioning as well as complex emotions/behaviors. The optogenetics technique due to its virtue of high spatial and temporal specificity, has uncovered neural correlates of organism functioning and behavior at a cellular/sub-cellular resolution, something not possible before in Neuroscience. Using the SciVerse Scopus database, we analyzed neuroscience based published literature (i.e., articles and reviews) utilizing Optogenetics as an investigative tool between years 2010 to 2020. 6568 published documents were refined to 2621 documents after using the appropriate filters in Scopus. The average retrieved document count per year was 262, with an over all *h*-index at 128. Karl Deisseroth and Edward S. Boyden were the two leading authors in the neuroscience optogenetics domain. The highly used author keywords in optogenetics were pertaining to *in vivo* models used, cell types and brain regions studied, etc. Top-cited documents in optogenetics were focused on optogenetics as a versatile developing tool, and its applicability as an investigative tool in diverse neuroscience domains. Thus, the increasing volume of published literature utilizing optogenetics signifies its enormous contribution as a discovery-based tool.

Keywords: Optogenetics, Neuron excitation, Neuron inhibition, Circuits, Behavior, VOSviewer.

INTRODUCTION

Our initial understanding of the involvement of the Central Nervous System (CNS) regions and circuits in functional behavioral output mainly rested on crude techniques such as electrode stimulation of brain areas, brain lesion studies, and pharmacological interventions.^[1-4] However, the non-specificity of these interventions made deciphering specific neurons and circuits involved in the particular investigated behavior among heterogeneous populations difficult to pinpoint. Thus, assigning the activity of specific neuronal populations with heterogeneous brain regions to be the underlying cause of distinct normal and abnormal physiology or behaviors became complicated and frustrating.

Electrophysiological recording in brain slices and *in vivo*, along with pharmacological approaches, did allow for a dissection of neural circuitry related to functional outcomes.^[5-7] Further more, technology to implant multichannel electrodes in brain



DOI: 10.5530/jscires.12.1.001

Copyright Information : Copyright Author (s) 2023 Distributed under Creative Commons CC-BY 4.0

Publishing Partner : EManuscript Tech. [www.emanuscript.in]

Correspondence: Dwight Figueiredo

Symbiosis Institute of Health Sciences (SIHS), A Constituent of Symbiosis International (Deemed University) (SIU), Pune-412115, Maharashtra, INDIA. Email id: dwight.fig@gmail.com ORCID ID: 0000-0003-4548-1998

Received: 13-06-2022; Revised: 10-12-2022; Accepted: 16-02-2023.

regions evolved and activity at the level of single neurons could be obtained.^[8-11] However, gaining stable high-quality recordings of single unit activity in certain hard-to-interrogate brain regions in awake moving animals while they carried out behavioral tasks proved to be technically difficult.^[12,13] Moreover, the single units recorded are sometimes difficult to decode given the heterogeneity of distinct neuronal populations in those recorded regions.^[14,15]

Genetically encoded calcium indicators with high regional and cellular specificity did help with neuronal activity monitoring at the population as well as to some extent at single cell resolution. ^[16-18] However, these technologies are not able to manipulate the activity of genetically defined cell types within heterogeneous brain regions with a level of spatial and temporal precision that could identify cause-effect relationships.

With the rapid development and coming together of three independent and mutually exclusive areas, 1. Applicability of light sensitive bacterial Rhodopsins to eukaryotic systems. 2. Non-genetic and genetic based *in vitro* and *in vivo* targeting strategies, and 3. The optical neural interface technology required for light delivery and recording *in vivo* optogenetics has become a mainstay technique in neuroscience to unravel the causal and associated neural underpinnings of physiological functioning, behavior, and CNS diseases (Table 1 for key publications, first author names, journals, year of publication, and times cited in these three complementary areas). The advantage of optogenetics compared to other techniques is its superiority in terms of cellular specificity and stimulation/inhibition temporal control (i.e., genetically defined cell population at millisecond level precision).

By year 2010, optogenetics was well established as an important technique in neuroscience which could be used to answer a multitude of hypothesis and question.^[19-21] Originating from the lab of Karl Deisseroth at Standford (in 2005), optogenetics spread to neuroscience labs in the United States and all across the globe between 2005 to 2010.^[22-24] Many neuroscience investigators have used optogenetics as a technique to answer challenging questions in their areas of specialization that were not adequately addressable before the advent of optogenetics.^[25-27] Keeping this in mind, we wanted to conduct a bibliometric analysis of neuroscience literature utilizing optogenetics as a technique. Utilizing bibliometrics, we wish to address the following objectives: i) Identify the most productive and impactful authors in optogenetics, ii) Identify author collaborative networks, iii) Conduct a citation analysis of literature and highlight important contributing publications by means of number of citations, and iv) Ascertain the most productive and collaborative journals, institutions, and countries. Towards this end, we conducted a bibliometric analysis on optogenetic based literature (i.e. articles and reviews) in neuroscience using the Scopus SciVerse database. With the ever-increasing use of optogenetics in neuroscience and other non-neuroscience fields, along with yet-to-be discovered applications, optogenetics as a technique still has enormous potential with a long way to go.

METHODOLOGY

Bibliographic database

In this study, Scopus was the database that was used due to its vast scientific, technical, medical and social sciences coverage.^[28] It being the largest database and its easy-to-use analytic functions made it a database of choice.^[29] The choices of filters used are given in supplementary Figure 1. In our analysis, we chose to include only journal articles and journal reviews, as these types of documents contains majority of the scientific findings and literature in the optogenetics area. This is the case since optogenetics is a technique applicable to many a neuroscience research question.

Search strategy and validity

In order to obtain documents using optogenetics in a neuroscience-based context, we used the following search string in Scopus: "((optogenetics) AND ("Neuroscience" OR "Nervous system"OR "Brain" OR "Circuit*" OR "inhibit* neuron*" OR "excite* neuron*"))". Using this Search String in SCOPUS, we obtained 2621 documents. For validity of the above search strategy, authors manually checked 100 articles which were the

top 10 cited articles for each of the top 10 authors during the study period.

Besides having no false positive results in our manual analysis, all of the documents scanned contained optogenetics in a neuroscience related context indicating correct scope.

Data Analysis

Bibliometric indicators/parameters such as highest number of citations, country productivity, authorship, citation analysis, annual growth, document type were presented as frequency and percentage. Annual Growth Rate (AGR), Relative Growth Rate (RGR), DT, citation count and characteristics, and quality of publications Hirsch index (*h*-index) were calculated and/or presented as in previous studies.^[30-33]

Collaboration and authorship analysis

Number of single authored articles and number of multi-authored joint articles were calculated in excel. Overall collaborations in the form of Degree of collaboration, Collaborative Index (CI), Average number of Authors Per Paper (AAPP) were calculated and presented as before.^[32,33]

Visualization and mapping

VOSviewer program was used to visualize bibliometric networks of author collaborations, country collaborations, institutional collaborations, Journal co-citation collaborative analysis,^[34,35] Network visualization was chosen as the mode of presentation, with color (one cluster or group of close units), circle size (greater productivity or citations), font size, and thickness of connecting lines strength of collaboration) representing distinct parameters in the map. In maps, higher the relative link strength suggested stronger collaborative ties between units. Regarding pictorial depiction of geographical distribution of publications, iMapBuilder Interactive HTML5 Map Builder software was used.

Statistical analysis and ethics

Descriptive statistics were presented as frequencies and percentages. Mean, median, and range were presented for continuous data; the interquartile range (Q1-Q3) was presented in case of medians.

Microsoft Excel was used for data analysis and presentation. Since this study did not include any human data nor human subjects, and was based solely on electronic data, this study was exempted from ethical approval.

RESULTS

Description of retrieved literature

Making use of the filter tools in Scopus we selected research articles and reviews utilizing optogenetics in neuroscience between years 2010-2020. Based on the given search string and

Figueiredo and Sanghavi.: Optogenetics as a Tool in Neuroscience: Bibliometric Mapping (2010-2020)

| Microbial Opsins | | | | | |
|---|--|----------------|--|--|--|
| Paper Highlights | First Author/Journal/Year | Times | | | |
| | | Cited | | | |
| Rhodopsin-like Protein: Halobacterium halobium | (Dieter Oesterhelt, Nature New Biology, 1971) | 1576 | | | |
| Rhodopsin-regulated calcium currents: <i>Chlamydomonas</i> | (Hartmann Harz, <i>Nature</i> , 1991) | 188 | | | |
| Bacteriorhodopsin expressed in <i>S. pombe</i> | (Hildebrandt V, Proc. Natl. Acad. Sci. U.S.A., 1993) | 24 | | | |
| Functional expression of bacteriorhodopsin in oocytes | (Nagel G, FEBS Lett., 1995) | 66 | | | |
| Two rhodopsins: Chlamydomonas reinhardtii. | (Sineshchekov OA, Proc. Natl. Acad. Sci. U.S.A., 2002) | 408 | | | |
| Channelrhodopsin-1 (ChR1): A light-gated proton channel in green algae | (Nagel G, <i>Science</i> , 2002) | 753 | | | |
| Channelrhodopsin-2 (ChR2): A directly light-gated cation-selective membrane channel | (Nagel G, Proc. Natl. Acad. Sci. U.S.A., 2003) | 1788 | | | |
| Red-shifted optogenetic excitation: Volvox carteri | (Zhang F, Nat. Neurosci., 2008) | 392 | | | |
| eNpHR: A Natronomonas halorhodopsin enhanced for optogenetic applications | (Viviane Gradinaru, Brain Cell Biol., 2008) | 350 | | | |
| Halorhodopsin: Improved expression and targeting | (Shengli Zhao, Brain Cell Biol., 2008) | 145 | | | |
| Channelrhodopsin-1 (ChR1) and-2 (ChR2): Molecular determinants | (Hongxia Wang, J. Biol. Chem., 2009) | 139 | | | |
| Engineered channel rhodopsin variants: Improved properties and kinetics | (John Y Lin, <i>Biophys. J.</i> , 2009) | 479 | | | |
| Bi-stable channel rhodopsins: Modification at the C128 position | (André Berndt, Nat. Neurosci., 2009) | 418 | | | |
| Channelrhodopsin-2: Structural guidance of the photocycle | (Christian Bamann, Biochemistry, 2010) | 163 | | | |
| Opsin expression | n-targeting genetic strategies | | | | |
| Paper Highlights | First Author | Times Cited | | | |
| Optical control of neural activity: Millisecond-timescale neural regulation | (Edward S Boyden, Nat. Neurosci., 2005) | 3087 | | | |
| Channelrhodopsin-2: Optical control of excitable cells | (Feng Zhang, Nat. Methods, 2006) | 532 | | | |
| Channelrhodopsin-2 Transgenic mice: <i>In vivo</i> light-induced activation of neural circuitry | (Benjamin R Arenkiel, Neuron, 2007) | 551 | | | |
| Channelrhodopsin-2 transgenic mice: High-speed mapping of synaptic connectivity | (H. Wang, Proc. Natl. Acad. Sci. U.S.A., 2007) | 304 | | | |
| A FLEX switch targets channelrhodopsin-2 to multiple cell types | (Deniz Atasov, J. Neurosci., 2008) | 471 | | | |
| Parvalbumin neurons and gamma rhythms enhance cortical circuit performance | (Vikaas S Sohal, C5 2009) | 1630 | | | |
| Phasic firing in dopaminergic neurons is sufficient for behavioral conditioning | (Hsing-Chen Tsai, Science, 2009) | 838 | | | |
| Activation of groups of excitatory neurons in spinal cord or hindbrain: locomotion | (Martin Hagglund Martin, Nat. Neurosci., 2010) | 190 | | | |
| Technological a | spects: In vivo/ Invitro optics | | | | |
| Paper Highlights | First Author | Times Cited | | | |
| Next-Generation Optical Technologies: Illuminating genetically targeted brain circuits | (Karl Deisseroth, J. Neurosci., 2006) | 512 | | | |

Table 1: Key initial publications involved in making optogenetics a mainstay technique in Neuroscience.

| Microbial Opsins | | | | | |
|--|--|------|--|--|--|
| Targeting and Readout Strategies for Fast Optical Neural Control: <i>In vitro</i> and <i>in vivo</i> | (Viviana Gradinaru, J. Neurosci., 2007) | 372 | | | |
| Optogenetic control of hypocretin neurons: Neural substrates of awakening probed | (Adamantidis AR, Nature, 2007) | 894 | | | |
| An optical neural interface: in vivo control of rodent motor cortex | (Alexander M Aravanis, J. Neural Eng., 2007) | 729 | | | |
| Sparse optical microstimulation in barrel cortex: Driving learned behaviour in freely moving mice | (Daniel Huber, <i>Nature</i> , 2008) | 372 | | | |
| Optical deconstruction of parkinsonian neural circuitry | (Viviana Gradinaru, Science, 2009) | 1106 | | | |
| Optogenetic interrogation of neural circuits: technology for probing mammalian brain structures | (Feng Zhang, Nature Protocols, 2010) | 516 | | | |
| Closed-Loop and Activity-Guided Optogenetic Control | (Logan Grosnick, Neuron, 2015) | 231 | | | |

А

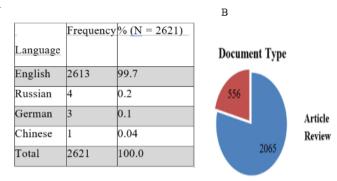


Figure 1: A) Document number based on medium of language. B) The number of articles and reviews included in the bibliometric analysis.

filters used 2621 documents were retrieved. English was the predominant language used in these retrieved publications (2615; 99.7%) (Figure 1A). Majority of these publications were articles (2065; 79%), and the rest were reviews (556; 21%) (Figure 1B).

A total of 1,05,381 citations were obtained during the study period having a mean of 40.2 ± 10.9 citations per document, a range of 0-157, and a median (Q1-Q3) of 5 (2–9) citations per document. The *h*-index of the retrieved documents stood at 131 (Table 2).

Mapping of author keywords using VOSviewer revealed 5 main clusters containing 217 author keywords such as transgenic mice, brain slice, somatostation, pyramidal cells, parvalbumin, cytology, interneuron, neural inhibition, gabaergic neurons, interneurons etc. in cluster 1 (Red-80 items); striate cortex, somatosensory cortex, cerebral cortex, brain cortex, visual stimulation, vision, wakefulness, etc. in cluster 2 (Green 71 items); anxiety, amygdala, avoidance behavior, basolateral amygdala, classical conditioning, dopamine, reward, motivation, nucleus accumbens, reinforcement, animal behavior, learning, memory, prefrontal cortex, etc. in cluster 3 (Blue-62 items); animal model, knockout mouse and pathology in cluster 4 (Yellow-3 items); and finally cell activation in cluster 5 (Purple-1 item) (Figure 2). Keywords with minimum occurrences of 50 times were shown in the map (217 keywords-5 clusters). Keywords having close spatial locations in the map have a higher chance of co-occurring together. For example, dopamine, dopaminergic nerve cell, motivation, reward, depression, odor, memory, learning, animal behavioral, anxiety, etc. occur close to each other in cluster 3 among 62 items signifying a close relationship and co-occurrence in documents (Note: "Optogenetics" with 1761 occurrences and a link strength of 39845 was removed from the analysis since it is the main component of the search term used).

Authorship pattern, collaboration, and prolific authors

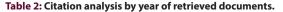
In terms of total number of publications, Deisseroth, K. (n = 59); Boyden, E.S. (n = 26); Yamanaka, A. (n = 17); Kravitz, A.V. (n = 16); and Lee, J.H. (n = 16) were the top 5 authors utilizing optogenetics as a technique in Neuroscience (Table 3). The current affiliation and geographical location of the institutions of these the authors are mentioned in Table 3.

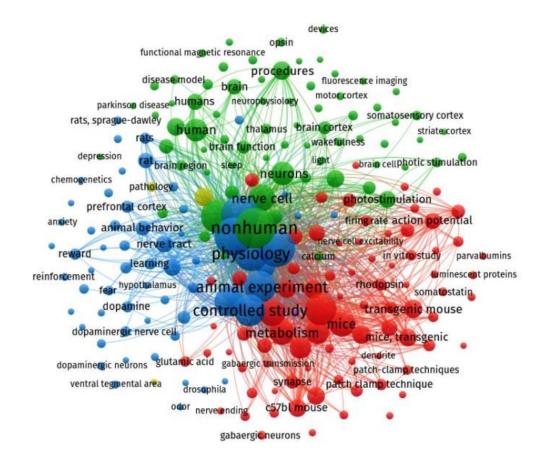
Using the VOSviewer software, top author collaborations in the field of optogenetics were mapped. The map included 56 authors-each represented by a circle-in 11 clusters (Figure 3).

In the collaborative author map (Figure 3), listed authors had a minimum of 10 publications and 100 citations. Adesnik H. and Lüthi A. were isolated as single author clusters whereas cluster 1, cluster 2 and cluster 3 had 11{chen x.; li b.; li h.; li x.; li y.; wang h.; wang l.; zhang j.; zhang x.; zhang y.; zhang z.}, 9{Augustine gj.; boyden e.s.; buzsaki g.; emiliani v,;

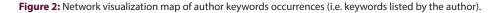
Feng g.; murphy t.h.; Svoboda k.; tye k.m.; zeng h.}, and 8{bonci a.; bruchas m.r.; de lecea l.; deisseroth k.; petersen c.c.h.; sohal v.s.; stuber g.d.; yizhar o.} authors respectively (Table 4). Deisseroth k. and Boyden e.s. are the top two authors with the highest total link strength signifying with these authors form different but connected hubs in the VOSviewer map (Figure 3; Table 4).

| Year | Frequency | % <i>n</i> = 2621 | Total Citations | Mean +/-SD of citations | Median (Q1-Q3) of citations |
|-------|-----------|-------------------|-----------------|----------------------------|-----------------------------|
| 2010 | 21 | 0.8 | 2610 | 124.3 ± 15.4 | 5 (1-12) |
| 2011 | 44 | 1.7 | 4264 | 96.9 ± 18.6 | 4 (1-11) |
| 2012 | 91 | 3.5 | 7447 | 81.8 ± 14.4 | 4 (2-10) |
| 2013 | 206 | 7.8 | 14138 | 68.6 ± 12.6 | 5 (2-11) |
| 2014 | 211 | 8 | 12217 | 57.9 ± 9.8 | 5 (2-11) |
| 2015 | 285 | 10.8 | 28240 | 99.1 ± 12.2 | 5 (2-10) |
| 2016 | 286 | 10.9 | 9989 | 34.9 ± 8.8 | 4 (1-9) |
| 2017 | 326 | 12.4 | 17080 | 52.4 ± 6.5 | 4 (1-9) |
| 2018 | 382 | 14.6 | 6141 | 16.1 ± 7.0 | 1 (0-6) |
| 2019 | 357 | 13.6 | 2575 | 7.2 ± 5.6 | 2 (0-5) |
| 2020 | 412 | 15.7 | 680 | 1.7 ± 2.4 | 1 (0-2) |
| Total | 2621 | 100 | 105381 | 40.2 ± 10.9 | 5 (2-9) |





A VOSviewer



Stronger research collaborations are indicated by closely packed circles. Deisseroth, K. was the author with the highest total number of Links (L = 20), Total Link Strength (TLS = 31) and documents (N = 57). This was followed by Boyden, E.S (L =

9; TLS = 12; N = 25); Kravitz, A.V. (L = 5; TLS = 10; N = 16); Yamanaka, A. (L = 3; TLS = 9; N = 17); and Lee, J.H.(L = 4; TLS = 9; N = 16) as the top five collaborative authors having highest total link strength (Table 5).

| Ranking | Author name | Total document count | Current Affiliation | Country |
|---------|----------------|----------------------|--|---------------|
| 1 | Deisseroth, K. | 59 | Stanford University, Palo Alto | United States |
| 2 | Boyden, E.S. | 26 | Massachusetts Institute of Technology, Cambridge | United States |
| 3 | Yamanaka, A. | 17 | Nagoya University, Nagoya | Japan |
| 4 | Kravitz, A.V. | 16 | Washington University School of Medicine in St. Louis, St. Louis | United States |
| 5 | Lee, J.H. | 16 | LVIS Corporation, Palo Alto | United States |
| 6 | Murphy, T.H. | 15 | The University of British Columbia, Vancouver | Canada |
| 7 | Nestler, E.J. | 15 | Nash Family Department of Neuroscience and Friedman Brain Institute, New York, NY | United States |
| 8 | Stuber, G.D. | 15 | University of Washington, Seattle | United States |
| 9 | Buzsaki, G. | 14 | NYU Langone Health, New York | United States |
| 10 | Kreitzer, A.C. | 14 | University of California, San Francisco, San Francisco | United States |

Table 3: Top 10 authors in the field of Optogenetics in Neuroscience, their current affiliations and locations.

The oldest authors related to the optogenetics field in terms of when they published their first document was BuzsÃiki, G. in 1973, Nestler, E.J. in1977, Murphy, T.H. in 1986, Deisseroth, K. in 1990, and Yamanaka, A. in 1997. Boyden, E.S.; Stuber, G.D.; Kreitzer, A.C. published their first document in 2000 which was followed by Lee, J.H. in 2002 and then Kravitz, A.V. in 2004 (Supplementary Table 1). The total publication count of Nestler, E.J. is the highest with 692 publications followed by Deisseroth, K. with 398 publications and BuzsÃ;ki, G. with 390 publications. Among the top listed authors in the optogenetics field Nestler, E.J.; Deisseroth, K.; and BuzsÃ;ki, G. have an h-index of 156, 135, and 132 respectively (Supplementary Table). In terms of total citation count, Nestler, E.J. and Deisseroth have a clear lead with 92273 and 68179 citations respectively over other authors in the field of optogenetics (Supplementary Table 1). The top cited article for each of the top 10 authors in optogenetics along with the year and journal of publication during the study time period is given in (Supplementary Table 1).

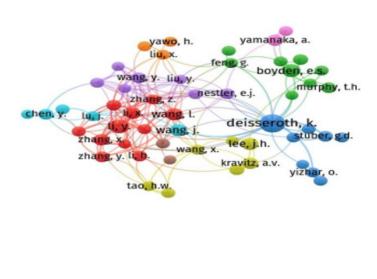
Top cited articles of the 10 most productive authors

The main article themes along with the key finding(s) and citation count of the top 10 cited articles for each of the top 10 authors are highlighted in Table 6. The top 10 cited articles were chosen for each author while reviews were excluded, as they do not contain original findings. The name of the first author, year

key points of article, and number of times cited are given in a supplementary excel document. The most cited articles for Deisseroth, K.(#1) within the study period include areas in optogenetic technological advancements (ChETA, Optetrodes), circuitry involved in depression, cognitive behaviors, aversion, countering seizures, reward-seeking behaviors and promotion of reward. Boyden, E.S.(#2) has his most cited in the optogenetics technological development sphere (Cre dependent optogenetic and intersectional targeting transgenic mice, high light sensitive optical neural silencer, Non-invasive optical inhibition-red shifted microbial rhodopsin, single cell resolution optogenetics) as well as circuitry involved in Slow Wave Sleep (SWS), primate visualmotor behavior, gamma and that a activity regulation, and optogenetic characterization of the temporal linearity of the MRI-BOLD response. Yamanaka, A.'s (#3) top 10 articles include articles related to top-down regulation of sensory perception, MCH neurons in sleep/wake regulation, glial based ischemic glutamate excitotoxicity, reversing pathological allodynia, NREM sleep regulation by VTA gabaergic neurons, aggressive behavior, top-down parasympathetic control, eNpHr2.0 Rosa 26 knock-in mice. Kravitz, A.V.s (# 4) top 10 cited articles include striatal circuitry associated with reinforcement, activity induced dsDNA breaks exacerbated by B-amyloid, basal ganglia gating, in dentification of striatal projection neuron subtypes, D1 versus D2 Medium Spiny Neuron (MSN) downstream pathway mapping,

of publication, title of article, journal, affiliation of first author,

adesnik, h.



lüthi, a.

Figure 3: VOSviewer map view of collaborations between top authors in optogenetics: Authors with a minimum of 10 publications and 100 citations were visualized. The map included 56 authors in 11 clusters who met the criteria of being active authors. Some names are not seen due to overlap of names. Clusters indicate active authors of close research collaboration.

cell specific interventions for motor recovery in dopamine deleted mice, constraining reward seeking, striatopallidal regulation of avoidance behavior, and medial amygdala pathway in experience-dependent aggression escalation.

Lee, J.H.'s (# 5) articles include central thalamus regulation of cortical and subcortical networks, hippocampal dorsal versus ventral downstream connectivity using opto-fMRI, opto-fMRI to validate a multivariate dynamic systems model for causal brain interactions, brain circuit function using dynamic causal modeling for Opto-fMRI, to study brain circuit function, seizure-like after discharge mapping using simultaneous LFP and opto-fMRI, *in vivo* human stem cell graft into host neural circuits, high throughput opto-fMRI imaging, and fMRI analysis methods for heterogeneous BOLD responses.

Murphy, T.H.'s (# 6) top cited articles included distinct cortical circuits for motor maps and complex forelimb movement, GABA neuron regulation of cortical blood flow, network dynamics monitoring post stroke, cortical output motor remapping and functional recovery post spinal injury, prostaglandin mediated E2 neurovascular coupling, set-up for chronic light-based motor mapping, PV neuron stimulation-evoked network activity recovery post global ischemia-somatosensory cortex, widespread deficits in motor output post stroke, resistance of

optogenetically evoked motor function to global ischemia and reperfusion, and mesoscale optical recordings to assess regional and functional connectivity. Nestler, E.J.'s (# 7) top cited articles included Antidepressant neuronal pathways-m PFC optogenetic stimulation, cocaine craving circuitry-Amygdala to NAc, PFC circuits in depression and anxiety-Cholecystokinin induced Delta Fos B, stimuli inducing Delta Fos B in MSNs, BDNF in mesolimbic reward pathway-gated by stress and CRF, chronic social-stress induced depressive behaviors, mapping brain metabolic connectivity with uPET and optogenetic stimulation, Mesolimbic BDNF mediating nociception in Chronic Neuropathic Pain, resilience to social stress in susceptible mice-alpha 1 and 2 Adrenergic pathways, and Orexin signaling in LHb modulating aggressive behavior in mice.

Stuber, G.D.'s (# 8) top cited articles include LH to LHb glutamatergic neurons regulating feeding and reward, medial preoptic area social reward circuit under hormonal control, LC to BLA noradrenergic anxiogenic circuit, inhibition of pre-synaptic GABA release in the BNST mediated by kappa opioid receptor signaling, vmPFC encoding of cocaine associated memory-temporal dynamics in recall and extinction, PVT projection neurons integrating signals for cue reward processing, inhibition of BLA to EC circuit disrupting acquisition of contextual fear, Local modulation of presynaptic release-DA terminals in the NAc, CeA neurotensin neurons in hedonistic behaviors, and optical suppression of drug-evoked phasic dopamine release. BuzsÃ;ki, G.'s (# 9) top cited articles include precise motor movement regulation-inhibition of purkinje cell activity, tracing corticocortical connections between motor forelimb areas, BG inputs inducing excitatory motor signals in the thalamus, stimulation of inputs in LA forms associative fear memory, probing neuronal populations-combining micromirror based optogenetics and VSDI imaging, NAc D2 MSNs in cocaine induced behavioral sensitization in-vivo excitatory GABA loop, improved luminopsins, and optogenetics as a tool for assessing functional synapse formation in neuronal connections in co-culture systems. Kreitzer, A.C.'s (# 10) top cited articles include dendritic versus perisomatic inhibition regulating activity of hippocampal place cells, micro-LEDs on silicon neural probes-high resolution optogenetic studies, circuitry underlying hippocampal ripple formation, induction of theta resonance in cortical circuits, spatio-temporal control of mutiple neurons in vivo-diode probes, implantable neural probes-monolithically integrated dieletric waveguide and recording electrodes, unraveling behavioral correlates of hippocampal and mossy cells, hippocampal place map stabilization by SPW-Rs, circuit architecture and dynamics in hippocampal network.

Table 6 The following data of the top 10 authors and their top 10 highest cited articles in descending order was retrieved from SciVerse Scopus with the given search string and filters-See supplementary file for more information for each published article. Reviews have not been considered, as these are not primary findings of an author, which this table intends to portray. Overlapping documents between authors, if any, have being mentioned only once with priority given to the more productive author in optogenetics)

Author collaborative clusters, citation clusters and highest cited document clusters

Neuroscience domain/area-wise VOSviewer constructed author collaborative and citation maps (top authors and highest cited documents) are displayed in Figure 4. The domains/areas in neuroscience represented in Figure 4 are domains in which optogenetics has made a significant contribution towards understand causal or associated circuitry. These represent the key research areas in neuroscience such as: Mood and Affective Behavior, Anxiety and Fear related Behaviors, Social behavioral states, Addictive and compulsive behavioral states, Motivational state (Reward versus risk), Cortical rhythms regulating Behavior, Learning and Memory, Epilepsy, Stroke and Neuro-degeneration. We ensured that authors that have significantly contributed to these areas (No. of publications in the area ≥ 3), and highly cited documents (No. of citations per document >= 50) are mapped using VOSviewer by setting appropriate thresholds. In any Vosviewer map, the size of the circles represent the weightage in the map, the thickness of the connecting links represents link strength, the distance between two spheres is inversely proportional to their closeness, and the units of a given cluster represent a closely connected group.

Preferred journals

Among the journals, the Journal of Neuroscience, Neuron, Nature Neuroscience and Elife were the preferred journals to publish articles using optogenetics as a technique. The journal of Neuroscience which is the official journal of the 'Society for Neuroscience' published 329(12.5%) documents, Neuron published 314 (12%) documents, Nature Neuroscience published 184 documents (7%), and Elife published 179 documents (6.8%) pertaining to Optogenetics.

Interestingly, these journals are top tier journals in the field of Neuroscience with Impact Factors (IF) 5.67 for 'Journal of Neuroscience', 14.415 for 'Neuron', 20.071 for 'Nature Neuroscience', 7.08 for 'Elife' and 3.156 for 'Frontiers in Neural Circuits'. This suggests that papers that include optogenetics as a technique to answer neuroscience-based research questions get published in high impact neuroscience journals. 'E-neuro' (2.9%), 'Journal of Neurophysiology' (2.9%), 'Frontiers in Cellular Neuroscience' (2.4%), 'Current Opinion In Neurobiology' (2.1%) and 'Journal of Visualized Experiments' (1,7%) are moderately popular with neuroscientists for publishing optogenetic related documents (Table 7). Using VOSviewer, a co-citation map of journals publishing a minimum of 500 citations in optogenetics was constructed. Based on the map, 42 journals were clustered in 4 distinct groups: cluster 1: 17 journals, cluster 2: 16 journals, cluster 3: 8 journals, cluster 4: 1 Journals (Figure 3). The 'Journal of Neuroscience' is most connected in terms of the number of lines it receives from other journals, indicating that this journal is most co-cited with other journals.

Furthermore, the journal of Neuroscience had the largest oval shape in the map indicating that this journal had the highest citation count in optogenetics (Figure 5).

In terms of total number of publications in the top 10 journals in the year 2020, '*Elife*' stood first at 5574, '*Journal of Visualized Experiments*' was in second with 5213 documents, and '*Journal of Neuroscience*' was third with 3561 documents (Supplementary Table 3). In keeping with the total publication count in 2020; '*Elife*' had the highest total citations count standing at 60, 463, followed by '*Journal of Neuroscience*' (37,553), '*Neuron*' (36,000), and '*Nature Neuroscience*' (25093) (Supplementary Table 3). The *h*-index of the top three journals in 2020 were 14, 21, and 22 respectively (Supplementary Table 3).

Interestingly articles relating to microglia and their functioning with a particular emphasis on signaling pathways associated with normal physiology and with disease conditions had the highest number of citations in the top three Neuroscience Journals in 2020. The articles titled, *"Microglia actively remodel adult hippocampal neurogenesis through the phagocytosis secretome"* in Journal of Neuroscience,

"TREM2 Regulates Microglial Cholesterol Metabolism upon Chronic Phagocytic Challenge" in Neuron, and "Lipid-droplet-accumulating microglia represent a dysfunctional and pro-inflammatory state in the aging brain" in Nature Neuroscience had citations counts of 51, 64 and 85 respectively in 2020 (Supplementary Table 4).

Among the top 10 articles in optogenetics (Table 8), five of them were reviews and five were articles. The Neuron Primer titled, "Optogenetics in Neural Systems" received the highest number of citations standing at 1157 which was published in Neuron in 2011. In this Neuron primer, OferYizhar from the karlDeisseroth group introduces reliable and targetable single component tools to excite (i.e. cation channels: Blue-green fast excitatory-ChR2, ChIEF, ChRGR, ChETAs etc.; Yellow-red fast excitatory-VChR1, C1V1 etc.), inhibit (Chloride pump: yellow-Red eNpHR3.0; proton pump: green-yellow Arch/ArchT, eBRetc), biochemically modulate (i.e., 500 nm: G-protein signaling-Opto-b2AR, Opto-a1AR, Rh-CT(5-HT1A); 450 nm: c-AMP signaling-bPAC, BlaC) and activate/deactivate through on/off switches (ChR2-step function opsins (SFOs), VChR1-SFOs). Besides the optogenetic channel tools, the primer talks about in vivo targeting of these optogenetic tools while achieving specificity through cell type specific viral promoters and Cre-driver mouse lines. This

| Cluster 1 | Cluster 2 | Cluster 3 | Cluster 4 | Cluster 5 | Cluster 6 | Cluster 7 | Cluster 8 | Cluster 9 | Cluster 10 | Cluster 11 |
|-----------|-----------------|---------------------|-------------------|------------------|-------------------|--------------|-------------------|-----------------|----------------|---------------|
| Chen, x. | Augustine, g.j. | Bonci, a. | kravitz, a.v. | chen, l. | chen, y. | liu, s. | arenkiel, b.r. | tanaka, k.f. | adesnik, h. | lüthi, a. |
| Li, b. | Boyden, e.s. | Bruchas, m.r. | kreitzer, a.c. | chen, z. | lu, j. | liu, x. | lu, y. | yamanaka, a. | - | - |
| Li, h. | Buzsaki, g. | de lecea, l. | lee, j.h. | liu, y. | surmeier, d.j. | yawo, h. | - | - | - | - |
| Li, x. | Emiliani, v, | deisseroth, k. | liu, j. | nestler, e.j. | wang, f. | - | - | - | - | - |
| Li, y. | Feng, g. | petersen, c.c.h. | tao, h.w. | wang, s. | wang, j. | - | - | - | - | - |
| Wang, h. | Murphy, t.h. | sohal, v.s. | wang, x. | wang, y. | - | - | - | - | - | - |
| Wang, l. | Svoboda, K. | stuber, g.d. | zhang, l.i. | zhang, s. | - | - | - | - | - | - |
| Zhang,j. | Tye, K.m. | yizhar, o. | - | - | - | - | - | - | | - |
| Zhang, x. | Zeng, h. | - | - | - | - | - | | - | | - |
| Zhang, y. | | - | - | | - | - | - | - | - | - |
| Zhang, z. | - | - | - | - | - | - | - | - | - | - |

flexibility in targeting particular types of cells in heterogeneous population of cells in a brain region allows investigators to ask and answer very specific questions. This document discusses certain technical difficulties and limitations, along with future challenges in the optogenetic field (i.e areas related to light propagation in brain tissue, two-photon laser setups and Implantable Fiber-optic Lightguides (IFL) etc.) The second most cited review focuses on reward circuitry in the brain and how it is perturbed in mood disorders like depression.

Evidence portraying depression related changes to reward related circuits in humans and mice/rats at the cellular and molecular level (i.e gene expression and epigenetic changes) is presented. These maladaptive structural and molecular changes lead to depression induced synaptic plasticity. However of note is that depression susceptibility varies based on genetics with many studies hinting at genes involved in susceptibility to depression or resilience. Interestingly, this review highlights the fact that depression and reward/addictive circuitry are linked and overlap at key nodes. Thus, it is conceivable that changes in such linked circuitry lead to lack of pleasure/reward seeking and/or dulled value perception of reward which is associated with depression. The third most cited article in optogenetics generates and characterizes four Cre dependent knock-in genetically engineered mouse lines with Channel Rhodopsins (i.e., Ai27: ChR2H134R-tdTomato; Ai32: ChR2H134R-EYFP), halorhodopsin (Ai39: eNpHR3.0-EYFP) and Archaerhodopsin (Ai35: ss-Arch-EGFP-ER2). The creation of these cre dependent optogenetic reporter lines increased the feasibility and usability of optogenetics in vivo given the vast choice of readily available Cre driver lines. For example, investigators could pick cells types

in the brain that they wished to optogenetically manipulate by crossing these reporter lines with a suitable Cre Driver line. The additional fluorescent labeling of opsin expressing cells visible and thereby easy to verify, experimentally manipulate, and observe in experimental setups. The review focusing on the neuronal circuits of fear and anxiety highlighted the achievements of optogenetics in identifying key neuronal circuitry associated with fear and anxiety. Optogenetics offers investigators spatial and temporal precision like never before thereby tremendously speeding up the discovery of circuitry related to fear and anxiety. Such reliable and precise optogenetic studies point to brain-wide neuronal networks connected by range projection pathways underlying fear and anxiety with local disinhibitory circuit gating as a common theme. Optogenetic studies show distinct but partially overlapping circuits involved in fear versus anxiety (e.g., Fear acquisition and expression: Basolateral Amygdala (BLA), central amygdala and the prelimbic region of the medial prefrontal cortex; anxiety: bed nucleus of the striaterminalis, the lateral septum, the Ventral Tegmental Area (VTA) and the BLA). Interestingly, optogenetic studies have shown that fear extinction involves distinct pathways in the same regions responsible for fear acquisition and expression, thereby implying that possible active cross-talk between these circuits may regulate the mechanism by which fear could be extinguished. Optogenetics allowed for detailed description of inhibitory neuronal type specific local afferent and efferent functional circuitry in the visual cortex. A combination of optogenetics and mapping techniques identified how different major subtypes of inhibitory neurons (i.e., parvalbumin, somatostatin and vasoactive intestinal peptide) talk to one-another and the output from such computations. Parvalbumin neurons strongly inhibit one another but don't connect much with

| ······································ | | | | | |
|--|---------------|------------|-----------|------------------------------|---------------|
| Author Ranking | Author name | Cluster No | Links (L) | Total Link Strength (TLS) | Documents (N) |
| 1 | Deisseroth K. | 3 | 20 | 31 | 57 |
| 2 | Boyden E.S. | 2 | 9 | 12 | 25 |
| 3 | Yamanaka A. | 9 | 3 | 9 | 17 |
| 4 | Kravitz A.V. | 4 | 5 | 10 | 16 |
| 5 | Lee J.H. | 4 | 4 | 9 | 20 |
| 6 | Murphy T.H. | 2 | 1 | 1 | 14 |
| 7 | Nestler E.J. | 5 | 6 | 9 | 15 |
| 8 | Stuber G.D. | 3 | 4 | 8 | 15 |
| 9 | Buzsaki G. | 2 | 3 | 3 | 14 |
| 10 | Kreitzer A.C. | 4 | 3 | 9 | 14 |

other neurons. In contrast, somatostatin interneurons inhibit all other interneuronal populations but not other somatostatin interneurons. VIP neurons preferentially inhibits somatostatin positive interneurons. In the article titled, "Evoked axonal oxytocin release in the central amygdala attenuates fear response" optogenetically stimulated hypothalamic oxytocin neuron terminals in the central Amygdala (CeA) caused fear suppression in fear conditioned rats. The lab from which optogenetics in neuronal systems originated-the Disseroth Lab-explained the scientific landscape over 10 years since its inception in the form of a historical Commentary titled, "Optogenetics: 10 years of microbial opsins in neuroscience." It details the conversion of optogenetics from a slow field possessing many technical hurdles for implementation to a rapidly growing and refined advanced neuronal perturbation technique (2010-2015), which has lead to many seminal discoveries of the underlying causal circuitry in certain functional outcomes and behaviors. The combination of optogenetics with previously discovered cell and tissue targeting strategies gave optogenetics an unmatched versatility. In the article by Alexxai V Kravitz et al., 2012 titled, "Distinct roles for direct and indirect pathway striatal neurons in reinforcement"; optogenetics was used to unravel specific effector pathways that are downstream of dopamine signaling. The authors show that preferential optogenetic activation of specific downstream dopamine pathways (i.e. D1 versus D2 pathways) could make the difference between induced persistent reinforcement or induced transient punishment. Thus, these circuits downstream of dopamine might be involved in encoding and expression of positive or negative valence associated with tasks that serve to guides future actions. The review by Kay M. Tye et al., 2012 titled," Optogenetic investigation of neural circuits underlying brain disease in animal models" highlights how optogenetics is the go to tool for delineating and obtaining a better understanding of underlying circuitry with a precision that was earlier not achievable earlier with the available techniques at that time. The topics covered in this review touched upon circuitry involved in

basic behaviors and disease states. Optogenetics played a major role in furthering our understanding of microcircuits in the amygdala underlying fear and anxiety, the hippocampus and its mechanisms of recall of remote memory, misbalanced Excitation/ Inhibition and rhythmic oscillations regulated by interneurons in autism and schizophrenia, circuits encoding reward relevant to addiction, and the basic circuitry involved in neurological disorders and possible treatment strategies. The article titled, "Ultrafast optogenetic control" provides evidence of the technical millisecond temporal precision of ChR2 light induced spikes. However, there are limitations in terms of ChR2 inducing gamma spike sustained trains (important physiological frequency band) and artificially generating optically induced plateau potentials of 10mv (i.e., up-states) that could make interpretation difficult. This summarizes the most-cited seminal literature in the optogenetics field between years 2010-2020.

Geographical distribution of publications and Institutions

The 15 most productive institutions using optogenetics as a technique are listed in Table 9. The top three institutions are Howard Hughes Medical Institute (135 documents; 5.2%), Massachusetts Institute of Technology (118 documents, 4.5%) and Stanford University (112 documents, 4.3%). Out of the top 15 institutes, eleven institutes are situated in the United States of America, three institutions from Europe, and one institution form China.

Network visualization map of institutional collaborations (Figure 6). Institutions having published a minimum of 25 documents are shown in the map (41 institutions, 6 clusters). Institutions with the same color were commonly listed together as collaborative institutes. So, for example, Riken center for brain science, University of Tokyo, Massachusetts Institute of Technology, and Japan Science and Technology agency have the same color suggestive that these institutions collaborate with each other in the optogenetics space.

Table 6: Highlighting top 10 cited documents by top 10 most productive authors.

| | # 1: Deisserot | h, Karl (6603640862) | |
|---|-----------------------------|---|----------------|
| Article Theme | Key Finding(s) | | Times Cited |
| Improved opsin: Ultrafast optogenetic control | ChETA. | | 496 |
| Antidepressant therapy | Activation of mP | Activation of mPFC. | |
| Gamma rhythm and cognitive behavioral impairment | Activation of NR | 1R deleted PV neurons. | 382 |
| Conditioned Place Aversion (CPA) | Activation of GA | BA expressing VTA neurons. | 364 |
| Interrupting Seizures | Closed-loop inhi cortex. | bition of thalamic neurons connected to the stroke injured | 355 |
| Primate optogenetics | Tools (promoters | , opsins, viral vector systems). | 337 |
| Increased motivation-based reward learning and behavioral flexibility | Increased phasic | DA neuron activity. | 236 |
| Optrodes development | Simultaneous in v | vivo optical activation and recording. | 233 |
| Reward Signaling | Glutamatergic V | ΓA to NAc signaling. | 219 |
| LHb output suppressed under rewarding conditions | Activation of VT. | A dopamine-GABA hybrid neurons targeting LHb pathway. | 210 |
| | # 2: Boyden, Edv | ward S. (35291447700) | |
| Article Theme | | Key Finding(s) | Times Cited |
| Cre Dependent Optogenetic transgenic mi | ce | Activation-(ChR2-td Tomato, ChR2-EYFP), Inhibtion- (halorhodopsin eNpHR3.0, archaerhodopsin Arch-ER2). | 731 |
| Intersectional mouse lines | | Cre/Flp and Cre/Dre for intersectional targeting of neuronal sensors/effectors. | 483 |
| Increased sensitivity opsin (Arch T) | | Three-fold increased sensitivity for neuronal inhibition over conventional Arch. | 319 |
| Red Shifted Opsin (JAWS) | | Three-fold more sensitivity to red light; Non-invasive transcranial optical inhibition induced. | 297 |
| Slow Wave Sleep Induction | | Silencing orexin/hypocretin hypothalamic neurons in inactive period. | 171 |
| Distinct Inhibitory regulation of gamma/th different time scales | neta rhythms on | Granule cells regulate gamma synchrony; glomerular cells regulate theta oscillations. | 100 |
| Temporally precise single-cell resolution o | ptogenetics | Two-photon holographic connectivity mapping in slices using soma targeted CoChR. | 99 |
| Defective Monkey Visuomotor behavior (i movements) | .e., Saccadic eye | Activation of Superior Colliculus neurons. | 98 |
| MRI BOLD optogenetic characterization | | Mouse opto-Fmri showed that optogenetically locally induced neuronal activity drives BOLD (increased activity confirmed electro physiologically). | 88 |

| Slow Wave Sleep Induction and Increased wakefulness to sleep transitions | Chronic inhibition of orexin neurons (Arch-T) during active phase. | | 83 | | | |
|--|--|-----|----|--|--|--|
| # 3: Yamanaka, Akihiro (55319368300) | | | | | | |
| Article Theme key Finding(s) | | | | | | |
| Accurate sensory perception | Top-Down Circuit M2-S1. | 169 | | | | |
| Sleep/wake regulation | Increased MCH activity-transition from NREM to REM sleep; MCH ablation-increased Wakefulness. | 131 | | | | |
| Ischemic excitotoxic glutamate induced brain damage | pH based regulation of astrocyte glutamate release. | 112 | | | | |
| Induction of slow wave sleep | Long lasting silencing of orexin/ hypocretin neurons. | 81 | | | | |
| State of wakefulness | Increased activity of LH orexin neurons. | 30 | | | | |
| Reversing pathological allodynia | Stimulation of VTA- NAc Pathway. | 30 | | | | |
| Regulation of NREM sleep | Increased VTA (GAD67)—LH (Orexin/ Hypocretin) promoted NREM sleep. | 21 | | | | |
| Aggressive behavior modulation | LH (Orexin)—LHb (GAD2); stimulation of LHb-GAD2 activity increases male-male aggression and reward in aggressive contexts. | 17 | | | | |
| Hypothalamic parasympathetic control of heart | Hypothalamic Orexin neurons control of cardiac vagal neurons in DMV. | 13 | | | | |
| Light induced neuronal silencing in knockin ROSA-26 mice | Microbial Halorhodopsin eNpHR 2.0. | 8 | | | | |

| # 4: Kravitz, Alexxai V. (6602759239) | | | | | |
|---|--|-------------|--|--|--|
| Article Theme | Key Finding(s) | Times Cited | | | |
| D1 versus D2 pathways in reinforcement learning | Activation of D1 neurons: persistent reinforcement; Activation of D2 neurons: transient punishment. | 566 | | | |
| Neuronal physiological activity leading to dsDNA breaks | Activity in Indirect pathway striatal neurons leads to expression of yH2A.X(i.e. DNA damage marker). | 259 | | | |
| Basal ganglia output regulation | Activation of D1 direct MSNs inhibit SNr neurons-movement initiation; Activation of indirect D2 MSNs-motor suppression. | 211 | | | |
| Optogenetic identification of striatal neurons | Distinct light-evoked spike properties (143 single units recorded-133 putative MSNs, 15 Putative FS interneurons). | 58 | | | |
| Brain wide dissection of D1 versus D2 MSN downstream pathways | Opposing BOLD Responses in downstream regions correlated with single unit activity. | 58 | | | |
| Motor recovery in dopamine deleted mice | Cell specific pallidal intervention (increased PV-GPe; decreased Lhx6-GPe activity rescues motor deficits). | 44 | | | |
| Constraining reward seeking | Global Ventral Pallidum (VP) activity increase induces robust place preference; increased glutamatergic activity in VP induces place avoidance. | 43 | | | |

| Feeding behavior | Increased activity of AgRP neurons in hypothalamus. | 24 | | | |
|--|--|--------------------|--|--|--|
| Approach-Avoidance behavior | Increased dorsolateral striato-pallidal iMSN neuron activity increase avoidance; decreased activity reduces avoidance. | 11 | | | |
| Aggression Modulation (Effect of traumatic stress on aggression) | Increased activity of MeApv-VmH and MeApv-BNST leads to aggression priming. | 5 | | | |
| # 5: Lee, Jin H | yung (57196135833) | | | | |
| Article Theme | Key Finding(s) | Times Cited | | | |
| Central thalamus arousal regulation | High frequency thalamic stimulation-arousal; whereas low frequency-behavioral arrest. | 68 | | | |
| Dorsal vs. Ventral hippocampal connectivity | Stimulation of Intermediate Hippocampus-widespread cortical and sub-cortical activation; Stimulation of Dorsal Hippocampus-activation restricted to hippocampus. | 50 | | | |
| Establishing causal brain interactions-Multivariate dynamic systems model | Optogenetic fMRI to validate computational causal interaction estimating methods. | 29 | | | |
| Dynamic causal modeling for systems level circuit interactions and Neuromodulatory therapies | Validated by Optogenetic fMRI data in basal ganglia-thalamocortical network. | 27 | | | |
| Seizure like after-discharges | Distinct recruitment of brain networks (measured by LFPs and fMRI) by after-discharges versus sub-threshold hippocampal stimulation using carbon based optrodes. | 25 | | | |
| Connectivity of human stem cell graft with host neural circuits | Novel Opto-fMRI imaging platform to study graft-host connectivity. | 21 | | | |
| Efficient real-time interactive high-throughput Opto-MRI imaging with parallel computation | 1.7% of TR. | 22 | | | |
| Opto-fMRI | Enabling causal tracing of activities arising from defined cell types and firing patterns across the whole brain. | 17 | | | |
| fMRI analysis methods for heterogeneous BOLD responses | General Linear Model (GLM) inferior to gamma basis set and Fourier basis sets. | 11 | | | |
| Opto-fMRI | Non-invasive method to measure global dynamic response (BOLD) to optogenetic stimulation of specific neural circuits. | 9 | | | |
| # 6:Murphy, Timothy H. 7401632487 | | | | | |
| Article Theme | Key Finding(s) | Times Cited | | | |
| Complex forelimb movements | Through intra-cortical synaptic transmission while topography of motor maps via segregated outputs. | 93 | | | |
| Regulation of blood flow by GABAergic neuron activity | Increased GABA neuronal activity leads to increased blood flow independent of net neuronal activity. | 59 | | | |

| Optical mapping of network dynamics post stroke | Network wide downscaling and recovery after stroke different for peri-infact and surrounding non-affected regions. | 58 |
|--|--|--------------------|
| Re-establishment of cortical motor maps after spinal cord injury | Pivotal role of minor dorsolateral corticospinal pathway in mediating spontaneous motor recovery. | 57 |
| Molecular mediators in neurovascular coupling (NVC) | COX2 Prostaglandin E2 regulates NVC through vasodilatory EP2, EP4 receptors. | 48 |
| Chronic light-based motor mapping | Light pulses of 2 mW produced well-defined stable maps; useful strategy to assess cortical reorganization or sensory-motor integration. | 25 |
| Prolonged Parvalbumin (PV) Network deficits in Somatosensory cortex post global Ischemia | PV neuron excitability reversed quicker than stimulation-evoked GABAergic synaptic network activity. | 21 |
| Assessment of excitability and motor output after Ministrokes | Wide-spread deficits in motor output despite recovery of cortical neuron excitability. | 19 |
| Differential sensitivity of sensory and motor systems to global Ischemia. | Motor output recovers faster than sensory systems. | 10 |
| Optical recordings to assess regional functional connectivity. | Channel rhodopsin and voltage-sensitive dye imaging. | 9 |
| # 7: Nestler, | Eric J. 7102006227 | |
| Article Theme | Key Finding(s) | Times Cited |
| Anti-depressent effect | Optogenetic stimulation of the medial prefrontal cortex. | 410 |
| | contex. | |
| Incubation of cocaine craving | Unsilencing of amygdala-accumbens synapses as withdrawal period increased. | 191 |
| Incubation of cocaine craving Blockade of Cholecystokinin (CCK) mediated social defeat stress(Δ FosB) | Unsilencing of amygdala-accumbens synapses as | 191 178 |
| Blockade of Cholecystokinin (CCK) mediated social defeat | Unsilencing of amygdala-accumbens synapses as withdrawal period increased. Stimulation of cortico-amygdala blocks anxiogenic effect; stimulation of cortico-accumbens blocks | |
| Blockade of Cholecystokinin (CCK) mediated social defeat stress(Δ FosB) | Unsilencing of amygdala-accumbens synapses as withdrawal period increased. Stimulation of cortico-amygdala blocks anxiogenic effect; stimulation of cortico-accumbens blocks social avoidance defects. By pharmacological, emotional and optogenetic stimuli (limbic brain regions to Nucleus | 178 |
| Blockade of Cholecystokinin (CCK) mediated social defeat stress(Δ FosB) Δ FosB induction in striatal medium spiny neurons CRF and Stress gate BDNF in Nucleus Accumbens (NAc)-A | Unsilencing of amygdala-accumbens synapses as withdrawal period increased. Stimulation of cortico-amygdala blocks anxiogenic effect; stimulation of cortico-accumbens blocks social avoidance defects. By pharmacological, emotional and optogenetic stimuli (limbic brain regions to Nucleus Accumbens). Stimulation of Mesolimbic pathway increases | 178 156 |
| Blockade of Cholecystokinin (CCK) mediated social defeat stress(Δ FosB) Δ FosB induction in striatal medium spiny neurons CRF and Stress gate BDNF in Nucleus Accumbens (NAc)-A stress context detecting function | Unsilencing of amygdala-accumbens synapses as withdrawal period increased. Stimulation of cortico-amygdala blocks anxiogenic effect; stimulation of cortico-accumbens blocks social avoidance defects. By pharmacological, emotional and optogenetic stimuli (limbic brain regions to Nucleus Accumbens). Stimulation of Mesolimbic pathway increases BDNF in socially stressed mice but not naïve mice. Essential role of mesolimbic (VTA-NAc) BDNF | 178 156 135 |

| Resilience to social stress | Stimulation of LC-VTA signaling reverses depression related behavior in susceptible mice dependent on α1 and B3A adrenergic receptors. | 18 |
|--|---|-------------|
| Male aggressive behavior | Increased stimulation of the LH (orexin)- LHb(GAD2) leads to inhibitory tone in LHb and male-male aggressive behavior. | 18 |
| #8: Stuber, G | Garret D. 700714214 | |
| Article Theme | Key Finding(s) | Times Cited |
| LH-LHb circuit for regulation feeding and Reward | Stimulation of glutamaertergic LHA-LHb leads to avoidance behaviors; Inhibition of LHA-LHb leads to real time place preference; LHA neurons bidirectionally regulate feeding. | 150 |
| Socially engaged reward circuitry | Stimulation of mPOA to VTA steroid responsive neurotensin expressing neurons promotes rewarding phenotypes, social approach and striatal dopamine release. | 119 |
| Anxiety-like behavior | Stimulation of LC-BLA Noradrenergic projection increases anxiety and aversive behaviors through B-adrenergicr receptors. | 101 |
| KOR (Kappa Opiod Receptors) role in CeA-BNST stress response | Long range GABAergic CeA-BNST connections inhibited by KOR activation. | 99 |
| VmPFC in extinction and recall of cocaine associated memory-reorganization over time | Stimulation of VmPFC facilitated extinction of remote memory; inhibition of VmPFC impaired recall of recent memory. | 56 |
| Reward seeking behavior | Disrupting Paraventricular Thalamus (PVT) coding of reward predicting cues leading to abnormal behavioral cue discrimination. | 50 |
| Contextual fear-memory | Inhibiting the BLA-EC circuit during acquisition lead to a significant lowering in freezing in context re-exposure. | 47 |
| Local modulation of synaptic release | Differential synaptic mechanisms of optogenetic versus electrical stimulation of dopamine release in the NAc. | 42 |
| Consumption of ethanol and palatable food-sweet fluids | Stimulation of the Central Amygdala (Neurotensin positive) to Parabranchial Nucleus leads to reinforcement and increase of ethanol/saccharin consumption in non-dependent animals. | 23 |
| Suppressing dopamine transients in drug associated behaviors | Inhibition of VTA NpHR expressing neurons reduced frequency of dopamine transients. | 15 |

| # 9: Augustine, C | George J. 55663195000 | |
|---|--|--------------------|
| Article Theme | Key Finding(s) | Times Cited |
| Precise control of movement kinematics | Suppression of spontaneous activity of Purkinje cells is sufficient for discrete control over movements. | 135 |
| Generation of new lines of optogenetic based transgenic mice | Enhanced halorhodopsin, better versions of channelrhodopsin, Florescent tags, and neuron specific promoters. | 48 |
| Optogenetic tracing of corticocortical connections invivo (Motor Forelimb areas) | Determining functional connectivity between RFA and CFA. | 39 |
| Basal Ganglia (BG) inhibitory input drives motor signals in thalamus | Stimulation of GP-BG input leads to surge of APs in VL neurons and post-inhibitory muscle contraction. | 38 |
| Associative fear memories | Stimulation of presynaptic input (main auditory brain regions) into Lateral Amygdala with accompanied foot shock was sufficient to serve as conditioned stimuli and increase freezing response. | 32 |
| Probing functions of neuronal populations | Micromirror based presynaptic stimulation(cerebellar Interneurons) with post synaptic VSD (purkinje neurons) imaging. | 25 |
| Cocaine induced behavioral sensitization | Stimulation of NAc D2R-MSNs attenuated expression of cocaine induced behavioral sensitization. | 20 |
| Excitatory GABA loop operating invivo | Stimulation of Parallel fibers(PF) leads to PF excitation via GABA A receptors in cerebellar molecular layer. | 18 |
| Luciferase opsin combinations for improved luminopsins | Genetically encoded means of manipulating neuronal activity via chemogenetics and optogenetics (LMO4 and iLMO4). | 17 |
| Approach for assessing formation of neuronal connections in hybrid co-culture systems | Stimulation of rat cortical neurons leads to increase in frequency of currents in IPSC derived human neurons. | 14 |
| # 10: Buzsaki, | Gyórgy 7006676856 | |
| Article Theme | Key Finding(s) | Times Cited |
| Dissecting perisomatic and dendritic inhibition of hippocampal place cell activity | Inhibiting SOM interneurons increased burst firing; Inhibiting PV interneurons shifted theta spikes to the trough phase. | 353 |
| Monolithical tool for cellular circuit level analysis with spatiotemporal precision in freely moving animals | Four shank silicon probes consisting of 12 $\mu LEDs$ and 32 recording sites. | 214 |
| Hippocampal ripple oscillations | Temporally precise local pyramidal cell-interneuron interactions support ripple generation. | 192 |
| Inhibition induced theta band spiking | Role of PV cells and Ih Channels in post inhibitory spiking of pyramidal cells. | 173 |

| Diode probes for spatiotemporal optical control of multiple | Concurrent circuit monitoring and multisite | 158 |
|--|---|-----|
| neurons | perturbations. | 100 |
| Neural probe with monolithically integrated electrodes and waveguide | Optical targeting of single neurons with minimal power and monitoring with single unit recordings. | 130 |
| Granule cell-mossy cell interactions for pattern separation | Granule cells-sparse firing and single place field; Mossy cells-more active, multiple place fields and remapped under similar conditions. | 104 |
| Hippocampal pyramidal cell-interneuron circuitry and functional dynamics | Interneuron architecture and dynamics: Divergent synaptic connections-synchrony between interneurons; synchrony of convergent presynaptic inputs boosted postsynaptic drive; Spike transmission modulated by spike timing of post-synaptic cell. | 72 |
| Stabilizing hippocampal spatial maps | Scrambling sharp-wave ripples of place cells during learning caused remapping and no updating of spatial information. | 71 |

In terms of country productivity, the United States of America has the highest productivity with 1715 documents (46.15%) (Table 10). The countries that have productivity above 100 documents are: Germany (249; 6.8%), Japan (195; 5.3%), China (193; 5.2%), United Kingdom (181; 4.9%), Canada (174; 4.7), France (141; 3.8) and Switzerland (113; 3.1).

A heat-map portraying this distribution is shown in Figure 7.

The dark maroon colour indicates the United States which is the major contributor of published Literature on optogenetics (>15% of total published documents). The lighter the shade the lower the contribution of documents from that country. The countries with no filled in color shows negligible or no contribution at all to published Literature in optogenetics (< 0.1%). Among the South Asian countries, Australia ranks first with 51 documents, followed by Singapore (27), India (15), New Zealand (14), Taiwan (10), Hong Kong (7), Thailand (3), Pakistan (2) and Bangladesh (1). In the African continent, South Africa is the only contributor to documents related to optogenetics (4). In South America, the top three productive countries are Mexico (16), Denmark (15), and Chile (10).

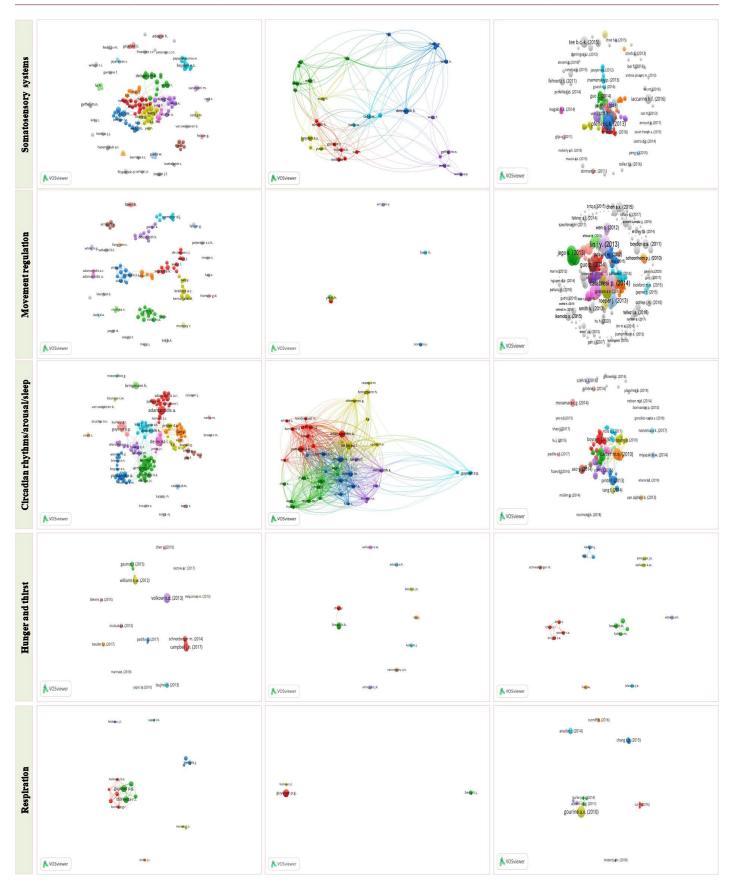
The thickness of connecting line between any two countries indicates strength of collaboration. For example, the link strength (collaboration) between USA and Germany was 84 and it represents a thick line. On the other hand, the line between USA and New Zealand had weak link strength of 1. Countries with similar color form one cluster. For example, countries with red color such as Spain, Austria, Hungary, Portugal, and Japan existed in one cluster and had the highest percentage of collaboration within this cluster (6 clusters; 24 items).

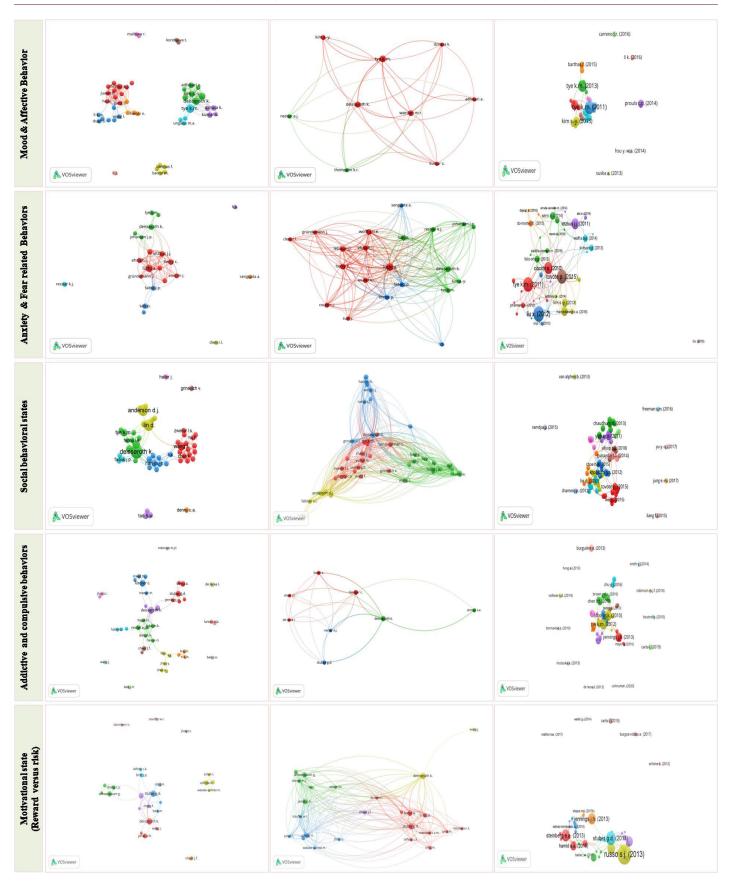
China, Canada, and Israel were clustered in yellow since the bulk of their collaboration is with the USA, so they are grouped with USA (Figure 8). In terms of cross-country collaborations, USA and germany had the highest link strength (i.e., 84), followed by USA and United Kingdom (i.e., 68), USA and China (i.e., 66), USA and Japan (i.e., 65), USA and Canada (i.e., 59). On the other hand, there are countries that were relatively spaced out in the map and had fewer thin connecting lines with other countries signifying weak collaborations between them (e.g., Argentina: Total Link Strength = 11, Documents = 14; and New Zealand Total Link Strength = 9, Documents = 11).

China, Canada, and Israel were clustered in yellow since the bulk of their collaboration is with the USA, so they are grouped with USA. In terms of cross-country collaborations, USA and germany had the highest link strength (i.e., 84), followed by USA and United Kingdom (i.e., 68), USA and China (i.e., 66), USA and Japan (i.e., 65), USA and Canada (i.e., 59). On the other hand, there are countries that were relatively spaced out in the map and had fewer thin connecting lines with other countries signifying weak collaborations between them (e.g. Argentina: Total Link Strength = 11, Documents = 14; and New Zealand Total Link Strength = 9, Documents = 11).

Growth of publications

The number of published documents applying optogenetics as a discovery-based tool increased year-on-year from year 2010-20 (Table 11). However, between the years 2010-13 there was a dramatic increase in the number of reviews and articles. During the years 2013-20, it increased gradually as is seen from the numbers. The year 2020 had the highest total number of documents with optogenetics (i.e. 412 documents) whereas 2010 had the lowest number of documents (i.e., 21 documents) (Table 11). A similar trend was seen in the number of citations that were obtained year-wise.





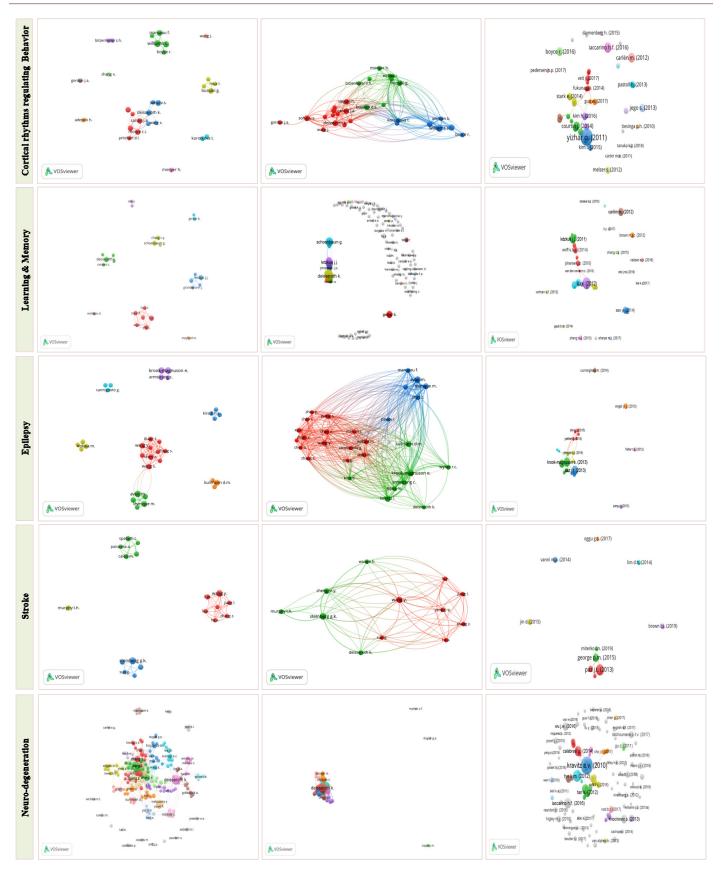


Figure 4: VOSviewer mapping based on top-author collaborations, top-citation clusters, and top-cited document clusters (i.e., documents consisting of articles and reviews) for different researched domains in Neuroscience using optogenetics: The left panel depicts author collaborative clusters, middle panel depicts citation based author clustering, and the right panel depicts citation based top-cited document clustering. Objects belonging to the same cluster in these maps are represented by the same color.

| ор | togenetics. | |
|---|------------------------|--------------------------------|
| Journal | Number of documents | Percentage of Documents (%) |
| Journal of Neuroscience | 329 | 12.6 |
| Neuron | 314 | 12.0 |
| Nature Neuroscience | 184 | 7.0 |
| Elife | 179 | 6.8 |
| Frontiers in Neural Circuits | 85 | 3.2 |
| Eneuro | 75 | 2.9 |
| Journal of Neurophysiology | 75 | 2.9 |
| Frontiers in Cellular Neuroscience | 62 | 2.4 |
| Current Opinion in Neurobiology | 55 | 2.1 |
| Journal of Visualized Experiments | 46 | 1.8 |
| Frontiers in Neuroscience | 45 | 1.7 |
| Cerebral Cortex | 44 | 1.7 |
| Journal of Neuroscience Methods | 44 | 1.7 |
| Neuroscience | 37 | 1.4 |
| Frontiers in Behavioral Neuroscience | 36 | 1.4 |
| Journal of Neural Engineering | 35 | 1.3 |
| European Journal of Neuroscience | 34 | 1.3 |
| Neurophotonics | 32 | 1.2 |

 Table 7: The journals that have published more than 30 documents in optogenetics.

The citation count almost doubled each year in the first few years post 2010, but then fluctuated between years 2013 to 2020 (Table 2). The highest citation count during the study period was in 2015 (i.e., 28240 citations); the lowest citation count was in the year 2020 (i.e., 680 citations). Interestingly, the number of citations was on the decline post 2018 (17080 in 2017 to 6141 in 2018 to 2575 in 2019 to 680 in 2020). The mean number of citations per document in general showed a decreasing trend between years 2010-20.

However, there was a precipitous drop between the years 2015 to 2016 (i.e., 99 to 35 citations per document) and between years 2017 to 2018 (i.e., 52 to 16 citations per document). The inter-quartile range (Q1-Q3) and the median number of citations for each year are mentioned in Table 2.

With regards to the statistics based on authors and authorship, the total number of authors increased from 86 in years 2010 to 3244 in year 2020 (Table 12). During the study period a total of

15937 authors contributed to this work (Table 12). There was an exponential increase in the number of authors contributing to documents in the initial years (i.e., 2010-2013), post which there was a steady increase in the number of documents (Table 12). The average number of authors per document increased from around 4 (2010) authors per document to around 8 authors per document (2020) (Table 12). This indicates that on an average the number of authors per document doubled during the study period (Table 12).

A study of growth rate indicated that the Average Growth Rate (AGR) fluctuated between-0.1 to 1.1 (Table 13). The Relative Growth Rate (RGR) declined from 1.1 to 0.2 during the study period (Table 13). The RGR was 1.1 in 2011, steadily decreased to 0.4 in 2015; and then steadied at 0.2 between years 2018-2020 (Table 13). An analysis of the Doubling Time (DT) revealed an increase from 0.6 in 2011 to 4.1 in 2020. There was an instance of a rapid increase in DT during the study period (i.e., between 2013 to 2014, DT values of 0.8 to 1.5). Since the values of RGR and DT were not stable during the study period, this indicated that the growth in publications was not exponential (Table 13). The number of single author publications fluctuated throughout the study period while the percentage of single author publications declined in general (2010: 6; 28.6-2020: 0; 0) (Table 14). As a corollary, the number and percentage of multi-author publications increased during the study period (2010: 15; 71.4-2020: 412; 100) (Table 14). The collaborative index increased from 5.3 in 2010 to 7.9 in 2020 (Table 14). The degree of research collaboration among the authors of papers using optogenetics was 92.8%.

DISCUSSION

Bibliometric indicators of literature pertaining to the technique 'optogenetics' in neuroscience were sought after in this study. Since the United States of America is the most productive country in terms of document count, it is but natural that the predominant language contained in these documents is English. Even though countries and institutions in Europe were substantial contributors, majority of the documents using optogenetics were published in English. One possibility could be that high impact neuroscience journals that publish optogenetic based literature accept articles only in English. This would be best to cater to their wide neuroscience readership base arising from many countries.

The total number of publications (i.e., publication count) as well as the total citation count increased over the study period which can be attributed to the increasing number of labs/and investigators incorporating optogenetics into their respective research hypothesis and questions, especially between 2010-15.^[21,36,37] The increasing total citation count might be attributed to the effectiveness of optogenetics as a potent discovery tool opening the doors to further important testable questions and hypothesis.^[39,40] Thus, future neuroscience based studies building upon these fundamental discovery based papers

| Rank | Title | Year | Journal name | Cited By | Type of document |
|------|---|------|--------------------------------|----------|------------------|
| 1 | Optogenetics in Neural Systems | 2011 | Neuron | 1157 | Review |
| 2 | The brain reward circuitry in mood disorders | 2013 | Nature Reviews Neuroscience | 861 | Review |
| 3 | A toolbox of Cre-dependent optogenetic transgenic mice for light-induced activation and silencing | 2012 | Nature Neuroscience | 690 | Article |
| 4 | Neuronal circuits for fear and anxiety | 2015 | Nature Reviews Neuroscience | 641 | Review |
| 5 | Inhibition of inhibition in visual cortex: The logic of connections between molecularly distinct interneurons | 2013 | Nature Neuroscience | 600 | Article |
| 6 | Evoked axonal oxytocin release in the central amygdala attenuates fear response | 2012 | Neuron | 561 | Article |
| 7 | Optogenetics: 10 years of microbial opsins in neuroscience | 2015 | Nature Neuroscience | 537 | Review |
| 8 | Distinct roles for direct and indirect pathway striatal neurons in reinforcement | 2012 | Nature Neuroscience | 533 | Article |
| 9 | Optogenetic investigation of neural circuits underlying brain disease in animal models | 2012 | Nature Reviews Neuroscience | 499 | Review |
| 10 | Ultrafast optogenetic control | 2010 | Nature Neuroscience | 480 | Article |



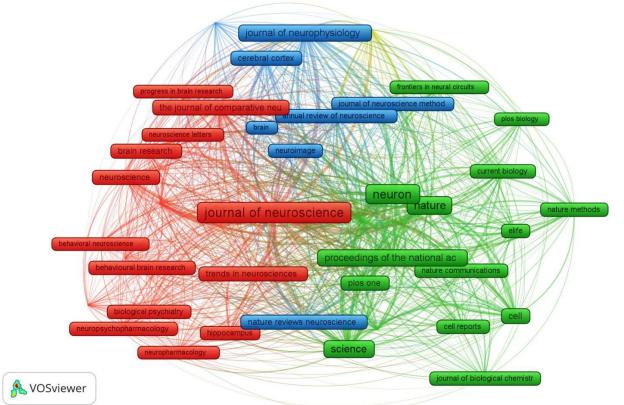


Figure 5: Network visualization map of journal co-citation analysis for journals which published documents in optogenetics with a minimum total of 500 citations. The Journal of Neuroscience one had many connecting lines with various journals indicating that this journal is being co-cited with various journals. Journals in the same cluster with the same color are being commonly co-cited together (42 journals; 4 clusters).

| lieuro | Jscience. | |
|--|----------------------------------|--------------------------------------|
| Affiliation | Number of Publications (N) | Percentage of Publications (%) |
| Howard Hughes Medical Institute | 135 | 5.2 |
| Massachusetts Institute of Technology | 118 | 4.5 |
| Stanford University | 112 | 4.3 |
| Harvard Medical School | 111 | 4.2 |
| CNRS Centre National de la Recherche Scientifique | 82 | 3.1 |
| National Institutes of Health NIH | 81 | 3.1 |
| Inserm | 78 | 3.0 |
| University of California, San Francisco | 73 | 2.8 |
| McGovern Institute | 72 | 2.7 |
| Columbia University | 71 | 2.7 |
| Chinese Academy of Sciences | 57 | 2.2 |
| University of Pittsburgh | 49 | 1.9 |
| University College London | 49 | 1.9 |
| Howard Hughes Medical Institute Janelia Farm Research Campus | 47 | 1.8 |
| University of California, Berkeley | 46 | 1.8 |

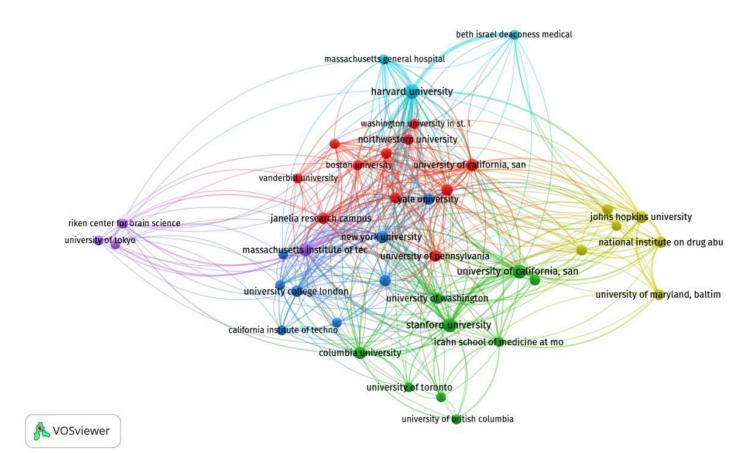
Table 9: The most productive institutes using optogenetics in neuroscience.

would lead to their citation count increasing over time. From year 2015-2020, the total number of citations has dropped even though the total number of publications has increased. This could be attributed to recent studies not having adequate time to be cited as well as the plateauing of the number of papers in the second half of the study period (i.e., drastic decrease in citation count per article).

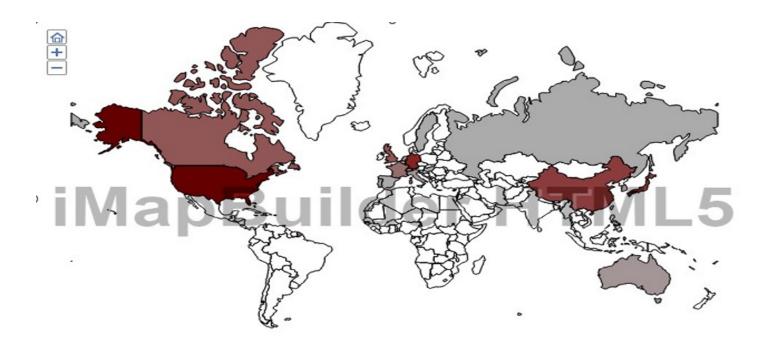
From the data, the average number of authors per document as well as the Collaborative Index (CI) kept on increasing throughout the course of the study period. This shows that optogenetics served as a favorable discovery-based tool for an increasing number of neuroscientists around the world. This was facilitated by the outstanding foundational contributions of Karl Diesseroth's and Ed Boyden's research clusters in the field of optogenetics.^[40-42] These research groups transformed optogenetics from a hard-toapply to a much easier and adaptable tool for neuroscience based research. Moreover, these discovered optogenetic tools were made easily available and widely accessible through viral repositories (e.g., Viral Vector Cores etc. for neurotrophic viruses containing opsingenes), mouse cores (e.g., Jax Lab for transgenic mice encoding opsin genes) as well as by training in the design, fabrication and usage of *in vivo* implantable optic fibers and probes (e.g., optrodes). Due to these initiatives promoting the dissemination of optogenetics around the globe, the total number of authors, the average number of authors per document, the degree of collaboration, and the collaborative index showed an increasing trend throughout the study period.

From data regarding the most productive authors and mapping of author clusters, Karl diesseroth is the most productive author with the most collaborative strength in the field of optogenetics. A second most productive author was his student Edward S. Boyden, having the second largest link strength. Not surprising, both authors were pioneers in the field of optogenetics providing evidence of the applicability of optogenetics as a tool in neuroscience in a seminal 2005 paper.^[40] More importantly, these two authors and their research clusters were instrumental in making optogenetics a mainstay technique in neuroscience for establishing casual underlying neural circuitry in behavioral/ physiological/disease states. However, one must not discredit the contributions of authors that formed the foundations for Eward S. Boyden's critical experiments in 2005, without which optogenetics in neuroscience would not have been possible.[43-49] Based on these game-changing findings by Eward S. Boyden and Karl Diesseroth, scientists of different nationalities and laboratories formed their research clusters and started to work together using optogenetics as a discover tool in their respective domains in neuroscience.^[50-52] In keeping with this geographical spread, we see clusters of American, Asian, and European authors which either directly or indirectly linked up with Ed Boyden and Karl diesseroth.^[21,23,53] In certain instances, scientists that were initially a part of the same research group in a particular lab using optogenetics, then moved on to start their own labs in different countries which contributed in a way to the spread of optogenetics as a technique across the globe.^[54-56]

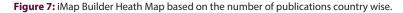
The USA ranked first in terms of number of neuroscience documents possessing optogenetics, in keeping with it being the country that houses most of the major institutions and authors contributing to optogenetic based neuroscience research (11 out of the 15 top institutions, and 8 out of the 10 top authors). Harvard Hughes Medical Institute (HHMI) is the major contributing institution signifying that optogenetics is used by an elite successful class of neuroscientists testing cutting edge ideas and hypotheses in Neuroscience. HHMI is followed by Massachusetts Institute of technology and Standford University in terms of institute productivity. In consonance with institutional productivity, Karl Deisseroth's lab is in Standford University, and Edward Boyden's lab is in Massachusetts Institute of Technology; both authors are currently HHMI funded. Interestingly, the HHMI funds scientists (based on their high quality of research and discovery), and not projects. With the money awarded to them, these scientists are thus able to spend the money on asking







Powered By iMapBuilder HTML5



| | Table To. The h | iost productive countries based on number of p | Jubications. |
|--------|-----------------|--|--------------------------------|
| S1. No | Country | No. of Publications | Percentage of Publications (%) |
| 1 | United States | 1715 | 46.5 |
| 2 | Germany | 249 | 6.8 |
| 3 | Japan | 195 | 5.3 |
| 4 | China | 193 | 5.2 |
| 5 | United Kingdom | 181 | 4.9 |
| 6 | Canada | 174 | 4.7 |
| 7 | France | 141 | 3.8 |
| 8 | Switzerland | 113 | 3.1 |



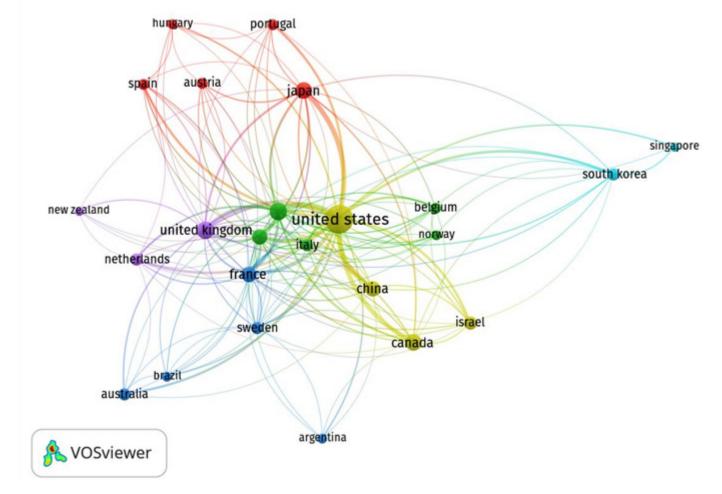


Figure 8: Network visualization map of international collaboration among countries with a minimum productivity of 10 documents (Minimum citations: 100).

and answering open-ended, difficult, and challenging questions. Thus, this lends credibility to the optogenetics technique, as it is used by well-renowned neuroscientists asking fundamental discovery-based questions related to Neuroscience and beyond.

In keeping with the high quality of neuroscientists, and of neuroscience using optogenetics as a technique, most of the publications using optogenetics were published in high impact neuroscience journals. Journals like Neuron, Nature Neuroscience, Journal of Neuroscience, Elife, and Frontiers in Neural Circuits. These journals have an Impact Factor (IF) greater than 5 suggesting high experimental rigor and stringent peer review of all included research documents. These journals publish original, novel, high-quality, authentic, cutting-edge, and supposedly

Table 11: The total number of document year wise (2010-2020).

| Year | No of Documents | Percentage of Documents (%) |
|-------|-----------------|-----------------------------|
| 2010 | 21 | 15.7 |
| 2011 | 44 | 13.6 |
| 2012 | 91 | 14.6 |
| 2013 | 206 | 12.4 |
| 2014 | 211 | 10.9 |
| 2015 | 285 | 10.9 |
| 2016 | 286 | 8.1 |
| 2017 | 326 | 7.9 |
| 2018 | 382 | 3.5 |
| 2019 | 357 | 1.7 |
| 2020 | 412 | 0.8 |
| Total | 2621 | 100 |

reproducible experimental research in the neuroscience domain. Thus, the content in these journals is deemed to be trustworthy by the neuroscience community and therefore have widespread readership in the USA, and around the globe.

European (i.e., Germany, United Kingdom, France, Switzerland, and Italy) and a few Asian countries (i.e., Japan, China, South Korea, Australia) have also contributed substantially in terms of published documents and this could be related to the better collaborative ties between institutes/labs in the United States and these countries. Moreover collaborative grants between labs/ investigators in these nations, and local funding sources in terms of grant agencies ensure the high productivity of high quality cutting edge research in Neuroscience. In keeping with this, the productivity of these countries is mirrored by the geographical positioning of grant agencies and bodies funding neuroscience applying optogenetics as a technique (i.e. National Institutes of Health, U.S. Department of Health and Human Services, National

Table 12: Average number of authors per document year wise (2010-2020).

| Year | Frequency | % (n= 2621) | Total number of Authors | Authors per document (Average) |
|-------|-----------|-------------|-------------------------|-----------------------------------|
| 2010 | 21 | 0.8 | 86 | 4.1 |
| 2011 | 44 | 1.7 | 163 | 3.7 |
| 2012 | 91 | 3.5 | 376 | 4.1 |
| 2013 | 206 | 7.9 | 1040 | 5.0 |
| 2014 | 211 | 8.1 | 1030 | 4.9 |
| 2015 | 285 | 10.9 | 1612 | 5.7 |
| 2016 | 286 | 10.9 | 1549 | 5.4 |
| 2017 | 326 | 12.4 | 1914 | 5.9 |
| 2018 | 382 | 14.6 | 2733 | 7.2 |
| 2019 | 357 | 13.6 | 2190 | 6.1 |
| 2020 | 412 | 15.7 | 3244 | 7.9 |
| Total | 2621 | 100 | 15937 | 6.1 |

Table 13: Average Growth Rate (AGR), Relative Growth Rate (RGR), and Doubling Time (DT) of documents year wise (2010-2020).

| Year | Frequency | AGR | % AGR | Cumulative total publications | loge W | RGR | DT |
|------|-----------|------|-------|-------------------------------|--------|-----|-----|
| 2010 | 21 | | | 21 | 3.0 | | |
| 2011 | 44 | 1.1 | 109.5 | 65 | 4.2 | 1.1 | 0.6 |
| 2012 | 91 | 1.1 | 106.8 | 156 | 5.0 | 0.9 | 0.8 |
| 2013 | 206 | 1.3 | 126.4 | 362 | 5.9 | 0.8 | 0.8 |
| 2014 | 211 | 0.0 | 2.4 | 573 | 6.4 | 0.5 | 1.5 |
| 2015 | 285 | 0.4 | 35.1 | 858 | 6.8 | 0.4 | 1.7 |
| 2016 | 286 | 0.0 | 0.4 | 1144 | 7.0 | 0.3 | 2.4 |
| 2017 | 326 | 0.1 | 14.0 | 1470 | 7.3 | 0.3 | 2.8 |
| 2018 | 382 | 0.2 | 17.2 | 1852 | 7.5 | 0.2 | 3.0 |
| 2019 | 357 | -0.1 | -6.5 | 2209 | 7.7 | 0.2 | 3.9 |
| 2020 | 412 | 0.2 | 15.4 | 2621 | 7.9 | 0.2 | 4.1 |

| | | Table 14: N | lumber of si | ngle-authored pub | lications, number | of multi-authored publi | Table 14: Number of single-authored publications, number of multi-authored publications, Degree of Collaboration and Collaborative Index (CI). | boration and Collabora | tive Index (Cl). | |
|-------|-----------|----------------|----------------------------------|--|---|---|--|---|----------------------------|-----------------------------|
| Year | Frequency | % (n= 4546) | Total number of Authors | No of Single Author publications | % Of Single Authored publications | Number of Multi-Authored Publications | % Of Multi-authored publications | Number of authors in multi-Authored publications | Degree of collaboration | Collaborative Index (Cl) |
| 2010 | 21 | 0.5 | 86 | 6 | 28.6 | 15 | 71.4 | 80 | 0.7 | 5.3 |
| 2011 | 44 | 1.0 | 163 | 8 | 18.2 | 36 | 81.8 | 155 | 0.8 | 4.3 |
| 2012 | 91 | 2.0 | 376 | 10 | 11.0 | 81 | 89.0 | 366 | 0.9 | 4.5 |
| 2013 | 206 | 4.5 | 1040 | 10 | 4.9 | 196 | 95.1 | 1030 | 1.0 | 5.3 |
| 2014 | 211 | 4.6 | 1030 | 8 | 3.8 | 203 | 96.2 | 1022 | 1.0 | 5.0 |
| 2015 | 285 | 6.3 | 1612 | 6 | 3.2 | 276 | 96.8 | 1603 | 1.0 | 5.8 |
| 2016 | 286 | 6.3 | 1549 | 10 | 3.5 | 276 | 96.5 | 1539 | 1.0 | 5.6 |
| 2017 | 326 | 7.2 | 1914 | 8 | 2.5 | 318 | 97.5 | 1906 | 1.0 | 6.0 |
| 2018 | 382 | 8.4 | 2733 | 0 | 0.0 | 382 | 100.0 | 2733 | 1.0 | 7.2 |
| 2019 | 357 | 7.9 | 2190 | 14 | 3.9 | 343 | 96.1 | 2176 | 1.0 | 6.3 |
| 2020 | 412 | 9.1 | 3244 | 0 | 0.0 | 412 | 100.0 | 3244 | 1.0 | 7.9 |
| Total | 2621 | 100 | 15937 | 83 | 3.2 | 2538 | 96.8 | 15854 | 0.9 | 6.2 |

Institute of Neurological Disorders and Stroke, National Institute of Mental Health based in the USA; the European Commission, European Research Council, Deutsche Forschungsgemeinschaft in Europe; and the Japan Society for the Promotion of Science, National Natural Science Foundation of China in Asia).

Besides monetary considerations, installing an optogenetic setup and making sure it works appropriately requires substantial technological know-how and expertise in optics as well as electrophysiology. Firstly, an electrophysiological rig or implantable electrodes that have the capability of recording currents from cells/ populations of cells in vivo, ex vivo, or in vitro, is essential for labs that use optogenetics. Secondly, for in vivo studies, knowledge and capability of conducting stereotaxic surgeries to get the lentiviruses/adenoviruses carrying the opsin genes into the appropriate brain regions and requisite neurons is a must. Thirdly, using optogenetics in vivo for assessing circuitry casual or associated with certain behaviours/functions in real time requires implantation of the optic fiber into that particular region of the brain and making sure the optic fiber is stably placed while the animal is awake and behaving. Only under circumstances in which the optogenetics set-up is accurate, precise, and reliable, can this technique be used to answer neuroscience-based questions. Thus, financial resources and technical limitations could be some of the reasons hindering the application and usage of optogenetics in neuroscience labs within developing countries-some countries within the Asian sub-continent, Africa and South America. To bridge this gap, there is need for 1. More active collaborations between neuroscience labs that possess the optogenetics technique in the developed world and labs in developing countries, and 2. Labs that have the resources and technical expertise to pass on this knowledge to labs that wish to setup optogenetics in-house, especially neuroscience labs in developing countries. Writing collaborative grants between optogenetics-ready and optogenetics-devoid labs could be one such solution to reduce the financial burden, improve sharing of technical knowledge, and increasing the pace and quality of research in developing countries.

Literature using optogenetics in Neuroscience covered a wide range of areas from its applications in understanding underlying circuitry causative or associated with anxiety, depression, addiction, and motivation, to understanding local micro-circuit within, and long-range circuit connectivity between brain regions. Optogenetics gave neuroscientists the unique opportunity of identifying cellular pathways and circuitry involved in behaviours/behavioral actions due to its high spatial and temporal resolution. Due to optogenetics, the cellular (i.e., engrams) and the molecular underpinnings of learning and memory could be unraveled.^[57,58] Answers to these questions eluded neuroscientists previously due to lack of an effective interrogative technique possessing high spatio-temporal resolution. The optogenetics technique has lead to a better understanding of connections and

0

circuitry within the brain necessary and/or sufficient for basic physiological functioning (hunger, thirst, circadian rhythms, sleep and awake states, etc.) as well as causative/underlying disease states (Parkinsons, Alzheimers, Amylotrophic Lateral Sclerosis, Fronto-Temporal Dementia, Stroke, Epilepsy, etc.).^[59-64]

Despite such rapid pre-clinical progress using optogenetics over that past decade (i.e., studies in mice, rats and lower primates), the applicability of optogenetics on a large-scale level in humans remains as a question mark-particularly with regards to its applications in the CNS (i.e., brain).^[65,66]

However, a few studies have shown the applicability of optogenetics in the human visual system, restoring sight in visually deficient people by transducing retinal ganglion cells with light sensitive opsins restoring vision.^[67-69] Manipulating the activity of neurons in the human brain, spinal cord or peripheral nerves is currently not possible due to its invasiveness with respect to light delivery into the brain, and the lack of suitable methods to get opsins into the appropriate neurons or sets of neurons safely.^[65,66] Even with the blossoming of non-invasive optogenetic techniques, optogenetics in humans has a long road ahead in terms of safety and efficacy testing.^[70-72] Thus, with the possibility of unlimited therapeutic opportunities by applying optogenetics in humans, we all hope that optogenetics one day will have its time.

Even though optogenetics has no direct applications in human brains and brain circuits as of today, the data gleaned from pre-clinical studies are being used to inspire pharmacological as we'll as-pharmacological techniques. As a non-pharmacological technique, Deep Brain Stimulation (DBS) along with other neuromodulatory techniques are currently used to treat different neurological diseases such as parkinsons, epilepsy, essential tremor, dystonia and obsessive compulsive disorders.^[73-77] Recent studies have hinted that neuromodulatory techniques, particularly DBS, can also be used as a potential treatment option for addictions, chronic pain, dementia, depression, traumatic brain injury, and neurodegenerative disorders such as Alzheimers, Huntington's disease, Multiple Sclerosis etc.^[78-85] However, the underlying circuitry, pathways, cell types and connections giving rise to behaviours, and their association with disease states is still an area of active investigation-optogenetics playing a useful role in such investigations.^[86,87] Knowledge gained from such preclinical studies could one day be used to target specific circuits in the human brain by DBS or other neuromodulatory techniques.^[88] Thus, neuromodulatory techniques (i.e., DBS) inspired by optogenetics holds great promise for improving symptoms and in some cases reverting the course of disease of patients suffering from certain neurological disorders.^[89-92] Thus, brain neuromodulatory techniques can be delivered to specific regions, cell types and/or axonal pathways to treat brain diseases with minimum possible side effects, unlike conventional drugs causing many unwanted side effects.

Besides informing brain neuromodulation, optogenetics holds therapeutic promise for pain relief and pleasure/hedonistic regulation (opto-morphs (MOPR), Kappa (KOPR), delta (DOPR), and Nociceptin Receptors (NOPR)), retinal sight restoration (Opto-mGluR6), and in optogenetic modulation of cell signaling-light sensitive cell surface receptors, transcription factors etc.^[93-97] Optogenetic based gene editing techniques (Photo-activatable Cas9), and optogenetic enhanced regenerative capacity (light based cell regeneration techniques) applications have also shown immense therapeutic promise.^[98-100]

Our study had a few limitations

First, the Scopus database includes only published literature and therefore this study could be biased towards published literature as opposed to grey literature, indexed journals, and language used in publications. Thus, there is a possibility of leaving out relevant studies that are not listed in the Scopus database. Second, due to the search query never being ideal in terms of formulation, false positives and false negatives can lead to non-relevant studies being included or removed. Thus, the total document count is to be expected with a certain margin of error; however, this should be small enough not to remain a significant factor. Third, it is important to keep in mind that the Scopus database is updated frequently with some journals periodically added or removed; this could therefore affect the current total count when the same search string is used. Third, the document count detected by Scopus could be lower than the actual count in instances when a single author or institution is represented by more than one name affecting the productivity of authors and institutions.

Besides these short comings, to our knowledge this is the first study to bibliometrically review neuroscience literature using optogenetics as a technique within the chosen study period.

CONCLUSION

Optogenetics has the potential to advance our understanding of the brain at circuit level detail in ways that were not possible before its discovery. Over the past 10 years (2010-2020); optogenetics, along with in vivo chemogenetics and muti-channel electrophysiology, have been cornerstone techniques in elucidating circuits and how they relate to behavior. This would not be possible without optogenetic's temporal and spatial precision. Credit to the authors that converted optogenetics from a nascent hard-to-use technique (between 2005-2010) to a highly applicable technique capable of solving a multitude of neuroscience based research questions and hypothesis (between 2010-2020). Furthermore, the future of optogenetics lays in how optogenetics can be adapted to cure and alleviate human disease burden, in particular brain diseases and mental disorders. Thus, we have reason to hope that optogenetics could switch gears with existing and upcoming non-invasive technologies to usher in a new era in neuroscience with a human therapy-centric focus.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

REFERENCES

- David HN, Sissaoui K, Abraini JH. Modulation of the locomotor responses induced by D1-like and D2-like dopamine receptor agonists and D-amphetamine by NMDA and Non-NMDA glutamate receptor agonists and antagonists in the core of the rat nucleus accumbens. Neuropharmacology. 2004;46(2):179-91.
- Inoue T, Li XB, Abekawa T, Kitaichi Y, Izumi T, Nakagawa S, et al. Selective serotonin reuptake inhibitor reduces conditioned fear through its effect in the amygdala. European Journal of Pharmacology. 2004;497(3):311-6.
- Meunier M, Bachevalier J, Mishkin M, Murray EA. Effects on visual recognition of combined and separate ablations of the entorhinal and perirhinal cortex in rhesus monkeys. The Journal of Neuroscience: The Official Journal of the Society for Neuroscience. 1993;13(12):5418-32. Available from: https://www.ncbi.nlm.nih.gov/ pubmed/8254384
- Talan MI, Engel BT. Effect of electrical stimulation of "rewarding" areas of the hypothalamus on habituation and dishabituation to repeated mild cold exposures in C57BL/6J mice. Physiology and Behavior. 1989;45(3):603-6.
- Fisahn A, Neddens J, Yan L, Buonanno A. Neuregulin-1 Modulates Hippocampal Gamma Oscillations: Implications for Schizophrenia. Cerebral Cortex. 2008;19(3):612-8.
- 6. Li B, Woo RS, Mei L, Malinow R. The Neuregulin-1 Receptor ErbB4 Controls Glutamatergic Synapse Maturation and Plasticity. Neuron. 2007;54(4):583-97.
- Woo RS, Li XM, Tao Y, Carpenter-Hyland E, Huang YZ, Weber J, et al. Neuregulin-1 Enhances Depolarization-induced GABA Release. Neuron. 2007;54(4):599-610.
- Bloms-Funke P, Gernert M, Ebert U, Löscher W. Extracellular single-unit recordings of piriform cortex neurons in rats: Influence of different types of anesthesia and characterization of neurons by pharmacological manipulation of serotonin receptors. Journal of Neuroscience Research. 1999;55(5):608-19.
- McNaughton BL, O'Keefe J, Barnes CA. The stereotrode: A new technique for simultaneous isolation of several single units in the central nervous system from multiple unit records. Journal of Neuroscience Methods. 1983;8(4):391-7. Available from: https://www.sciencedirect.com/science/article/pii/0165027083900973?via% 3Dihub
- Saeb-Parsy K, Dyball REJ. Defined Cell Groups in the Rat Suprachiasmatic Nucleus Have Different Day/Night Rhythms of Single-unit Activity *in vivo*. Journal of Biological Rhythms. 2003;18(1):26-42.
- Sanghera MK, Trulson ME, German DC. Electrophysiological properties of mouse dopamine neurons: *In vivo* and *in vitro* studies. Neuroscience. 1984;12(3):793-801.
- Chorev E, Epsztein J, Houweling AR, Lee AK, Brecht M. Electrophysiological recordings from behaving animals-going beyond spikes. Current Opinion in Neurobiology. 2009;19(5):513-9. Available from: https://www.sciencedirect.com/science/article/ab s/pii/S0959438809001111?casa_token=XwghT625 5qYAAAAA:XQZrknd_IPk-CliTjR1 yEZu04SSRImh9VyKq6rxyCD9KuDuJ2sj2AyOEN_pZGxhW6UsTujfxqw
- 13. Supèr H, Roelfsema PR. Chronic multiunit recordings in behaving animals: Advantages and limitations. Progress in Brain Research. 2005;147:263-82.
- Horn CC, Friedman MI. Detection of single unit activity from the rat vagus using cluster analysis of principal components. Journal of Neuroscience Methods. 2003;122(2):141-7.
- Rousche PJ, Petersen RS, Battiston S, Giannotta S, Diamond ME. Examination of the spatial and temporal distribution of sensory cortical activity using a 100-electrode array. Journal of Neuroscience Methods. 1999;90(1):57-66.
- 16. Grienberger C, Konnerth A. Imaging Calcium in Neurons. Neuron. 2012;73(5):862-85.

 Available
 from:
 https://ac.els-cdn.com/S0896627312001729/1-s2.

 0-S0896627312001729 main.pdf?_tid=50226b60-71e9-433b-adc3 12c87b489093

 &acdnat=1541321306_6d1672c64f6fe785b92e0ae9d5e5ca0d
 12c87b489093
 12c87b489093
- Lütcke H. Optical recording of neuronal activity with a genetically-encoded calcium indicator in anesthetized and freely moving mice. Frontiers in Neural Circuits. 2010:9.
- Patel AA, McAlinden N, Mathieson K, Sakata S. Simultaneous Electrophysiology and Fiber Photometry in Freely Behaving Mice. Frontiers in Neuroscience. 2020;14:148.
- Cardin JA, Carlén M, Meletis K, Knoblich U, Zhang F, Deisseroth K, et al. Targeted optogenetic stimulation and recording of neurons in vivo using cell-type-specific expression of Channelrhodopsin-2. Nature Protocols. 2010;5(2):247-54.
- Carter ME, Adamantidis A, Ohtsu H, Deisseroth K, De Lecea L. Sleep Homeostasis Modulates Hypocretin-Mediated Sleep-to-Wake Transitions. Journal of Neuroscience. 2009;29(35):10939-49.
- 21. Deisseroth K. Controlling the Brain with Light. Scientific American. 2010;303(5):48-55
- Ciocchi S, Herry C, Grenier F, Wolff SBE, Letzkus JJ, Vlachos I, *et al.* Encoding of conditioned fear in central amygdala inhibitory circuits. Nature. 2010;468(7321):277-82. Available from: https://www.nature.com/articles/nature09 559
- Haubensak W, Kunwar PS, Cai H, Ciocchi S, Wall NR, Ponnusamy R, et al. Genetic dissection of an amygdala microcircuit that gates conditioned fear. Nature. 2010;468(7321):270-6.

- Zorzos AN, Boyden ES, Fonstad CG. Multiwaveguide implantable probe for light delivery to sets of distributed brain targets. Optics Letters. 2010;35(24):4133.
- Ozden I, Wang J, Lu Y, May T, Lee J, Goo W, *et al*. A coaxial optrode as multifunction write-read probe for optogenetic studies in Non-human primates. Journal of Neuroscience Methods. 2013;219(1):142-54.
- Pisanello F, Sileo L, Oldenburg Ian A, Pisanello M, Martiradonna L, Assad John A, et al. Multipoint-emitting Optical Fibers for Spatially Addressable in vivo Optogenetics. Neuron. 2014;82(6):1245-54.
- Saunders A, Granger AJ, Sabatini BL. Corelease of acetylcholine and GABA from cholinergic forebrain neurons. Elife. 2015;4:e06412.
- 28. Elsevier BV. Scopus preview-Scopus-Welcome to Scopus. www.scopus.com. Availabl e from: https://www.scopus.com
- Falagas ME, Pitsouni El, Malietzis GA, Pappas G. Comparison of PubMed, Scopus, Web of Science, and Google Scholar: Strengths and Weaknesses. The FASEB Journal. 2008;22(2):338-42. Available from: https://faseb.onlinelibrary.wiley.com/doi/full/10. 1096/fj.07- 9492LSF
- Kumar R Santha, Kaliyaperumal K. Scientometric Analysis of Global Publication Output in Mobile Technology. DESIDOC Journal of Library and Information Technology. 2015;35(4):287-92.
- Santhakumar R. Mapping of Mobile Technology Publications: A Scientometric Approach, K. Kaliyaperumal. DESIDOC Journal of Library and Information Technology. 2014;34(4):298-303.
- Sweileh WM, Al-Jabi SW, Abu Taha AS, Zyoud SH, Anayah FMA, Sawalha AF. Bibliometric analysis of worldwide scientific literature in mobile-health: 2006-2016. BMC Medical Informatics and Decision Making. 2017;17(1):1-2.
- Zafrunnisha N, Pullareddy V. Authorship pattern and degree of collaboration in psychology. ALIS. 2009;56(4). Available from: http://nopr.niscpr.res.in/handle/1234 56789/7264
- Van Eck NJ, Waltman L. Software survey: VOSviewer, a computer program for bibliometric mapping. Scientometrics. 2009;84(2):523-38.
- Van Eck NJ, Waltman L. Visualizing Bibliometric Networks. Measuring Scholarly Impact. 2014;285-320.
- Tsunematsu T, Tabuchi S, Tanaka KF, Boyden ES, Tominaga M, Yamanaka A. Long-lasting silencing of orexin/hypocretin neurons using archaerhodopsin induces slow-wave sleep in mice. Behavioural Brain Research. 2013;255:64-74.
- Walsh JJ, Friedman AK, Sun H, Heller EA, Ku SM, Juarez B, *et al.* Stress and CRF gate neural activation of BDNF in the mesolimbic reward pathway. Nature Neuroscience. 2013;17(1):27-9.
- Boyden ES. A history of optogenetics: The development of tools for controlling brain circuits with light. F1000 Biology Reports. 2011;3.
- Deisseroth K. Optogenetics: Development and application. Neuroscience Research. 2009;65:S26.
- Boyden ES, Zhang F, Bamberg E, Nagel G, Deisseroth K. Millisecond-timescale, genetically targeted optical control of neural activity. Nature Neuroscience. 2005;8(9):1263-8.
- Adamantidis AR, Zhang F, Aravanis AM, Deisseroth K, De Lecea L. Neural substrates of awakening probed with optogenetic control of hypocretin neurons. Nature. 2007;450(7168):420-4. Available from: https://www.nature.com/articles/nature0631 0#citeas
- Aravanis AM, Wang LP, Zhang F, Meltzer LA, Mogri MZ, Schneider MB, et al. An optical neural interface: *In vivo* control of rodent motor cortex with integrated fiberoptic and optogenetic technology. Journal of Neural Engineering. 2007;4(3):5143-56.
- Harz H, Hegemann P. Rhodopsin-regulated calcium currents in Chlamydomonas. Nature. 1991;351(6326):489-91. Available from:
- 44. https://www.nature.com/articles/351489a0
- 45. Hildebrandt V, Fendler K, Heberle J, Hoffmann A, Bamberg E, Buldt G. Bacteriorhodopsin expressed in *Schizosaccharomyces* pombe pumps protons through the plasma membrane. Proceedings of the National Academy of Sciences. 1993;90(8):3578-82. Available from: https://www.pnas.org/content/pnas/90/8/3578 .full.pdf
- Nagel G, Möckel B, Büldt G, Bamberg E. Functional expression of bacteriorhodopsin in oocytes allows direct measurement of voltage dependence of light induced H+ pumping. FEBS Letters. 1995;377(2):263-6.
- Nagel G. Channelrhodopsin-1: A Light-Gated Proton Channel in Green Algae. Science. 2002;296(5577):2395-8.
- Nagel G, Szellas T, Huhn W, Kateriya S, Adeishvili N, Berthold P, et al. Channelrhodopsin-2, a directly light-gated cation-selective membrane channel. Proceedings of the National Academy of Sciences. 2003;100(24):13940-5. Available from: https://www.pnas.org/content/100/24/13940
- Oesterhelt D, Stoeckenius W. Rhodopsin-like Protein from the Purple Membrane of Halobacterium halobium. Nature New Biology. 1971;233(39):149-52. Available from: https://www.nature.com/articles/newbio233149a0.pdf
- Sineshchekov OA, Jung KH, Spudich JL. Two rhodopsins mediate phototaxis to low-and high-intensity light in *Chlamydomonas reinhardtii*. Proceedings of the National Academy of Sciences. 2002;99(13):8689-94. Available from: https://www.p nas.org/content/99/13/8689.full
- Chen Y, Lin YC, Kuo TW, Knight Zachary A. Sensory Detection of Food Rapidly Modulates Arcuate Feeding Circuits. Cell. 2015;160(5):829-41.

- 52. Liu D, Gu X, Zhu J, Zhang X, Han Z, Yan W, *et al*. Medial prefrontal activity during delay period contributes to learning of a working memory task. Science. 2014;346(6208):458-63.
- Stuber GD, Britt JP, Bonci A. Optogenetic Modulation of Neural Circuits that Underlie Reward Seeking. Biological Psychiatry. 2012;71(12):1061-7.
- English DF, Ibanez-Sandoval O, Stark E, Tecuapetla F, Buzsáki G, Deisseroth K, et al. GABAergic circuits mediate the reinforcement-related signals of striatal cholinergic interneurons. Nature Neuroscience. 2011;15(1):123-30.
- Diester I, Kaufman MT, Mogri M, Pashaie R, Goo W, Yizhar O, et al. An optogenetic toolbox designed for primates. Nature Neuroscience. 2011;14(3):387-97.
- Hardung S, Alyahyay M, Eriksson D, Diester I. A Toolbox for Optophysiological Experiments in Freely Moving Rats. Frontiers in Systems Neuroscience. 2017;11:27.
- Yizhar O, Fenno LE, Prigge M, Schneider F, Davidson TJ, O'Shea DJ, et al. Neocortical excitation/inhibition balance in information processing and social dysfunction. Nature. 2011;477(7363):171-8. Available from: https://www.ncbi.nlm.nih.gov/pmc/ articles/PMC4155501/
- Josselyn SA, Tonegawa S. Memory engrams: Recalling the past and imagining the future. Science. 2020;367(6473):eaaw4325. Available from: https://science.sciencem ag.org/content/367/6473/eaaw4325.abstract
- Liu X, Ramirez S, Pang PT, Puryear CB, Govindarajan A, Deisseroth K, et al. Optogenetic stimulation of a hippocampal engram activates fear memory recall. Nature. 2012;484(7394):381-5.
- Boyce R, Glasgow SD, Williams S, Adamantidis A. Causal evidence for the role of REM sleep theta rhythm in contextual memory consolidation. Science. 2016;352(6287):812-
- 61. Available from: http://science.sciencemag.org/content/352/6287/812.full
- Iaccarino HF, Singer AC, Martorell AJ, Rudenko A, Gao F, Gillingham TZ, et al. Gamma frequency entrainment attenuates amyloid load and modifies microglia. Nature. 2016;540(7632):230-5.
- Kong D, Tong Q, Ye C, Koda S, Fuller Patrick M, Krashes Michael J, et al. GABAergic RIP-Cre Neurons in the Arcuate Nucleus Selectively Regulate Energy Expenditure. Cell. 2012;151(3):645-57.
- Krook-Magnuson E, Armstrong C, Oijala M, Soltesz I. On-demand optogenetic control of spontaneous seizures in temporal lobe epilepsy. Nature Communications. 2013;4(1):1-8.
- Krook-Magnuson E, Szabo GG, Armstrong C, Oijala M, Soltesz I. Cerebellar Directed Optogenetic Intervention Inhibits Spontaneous Hippocampal Seizures in a Mouse Model of Temporal Lobe Epilepsy. Eneuro. 2014;1(1):ENEURO.0005-14.2014.
- Tye KM, Deisseroth K. Optogenetic investigation of neural circuits underlying brain disease in animal models. Nature Reviews Neuroscience. 2012;13(4):251-66.
- Shen Y, Campbell RE, Côté DC, Paquet ME. Challenges for Therapeutic Applications of Opsin-Based Optogenetic Tools in Humans. Frontiers in Neural Circuits. 2020;14:41.
- 68. White M, Mackay M, Whittaker RG. Taking Optogenetics into the Human Brain: Opportunities and Challenges in Clinical Trial Design. Open Access Journal of Clinical Trials. 2020;12:33-41. Available from: https://www.dovepress.com/ taking-optogeneticsinto-the-human-brain-opportunities-and-challenges-peer-reviewed-fulltext-article-OAJCT
- 69. Chapter 2-Restoring Vision to the Blind: Optogenetics. Translational Vision Science and Technology. 2014;3(7):4.
- Harris AR, Gilbert F. Restoring vision using optogenetics without being blind to the risks. Graefe's Archive for Clinical and Experimental Ophthalmology. 2021;260(1):41-5.
- Sahel JA, Boulanger-Scemama E, Pagot C, Arleo A, Galluppi F, Martel JN, et al. Partial recovery of visual function in a blind patient after optogenetic therapy. Nature Medicine. 2021;27(7):1223-9.
- Sun S, Shi J, Wang Y, Cheng J, Huang Z. A Temporal Precision Approach for Deep Transcranial Optogenetics with Non-invasive Surgery. Neuroscience Bulletin. 2021;37(8):1260-3.
- Tung JK, Shiu FH, Ding K, Gross RE. Chemically activated luminopsins allow optogenetic inhibition of distributed nodes in an epileptic network for Non-invasive and multi-site suppression of seizure activity. Neurobiology of Disease. 2018;109:1-10.
- Wang S, Kugelman T, Buch A, Herman M, Han Y, Karakatsani ME, et al. Non-invasive, Focused Ultrasound-facilitated Gene Delivery for Optogenetics. Scientific Reports. 2017;7(1):1-7.
- Bouwens Van Der Vlis TAM, Schijns OEMG, Schaper FLWVJ, Hoogland G, Kubben P, Wagner L, *et al*. Deep brain stimulation of the anterior nucleus of the thalamus for drug-resistant epilepsy. Neurosurgical Review. 2018;42(2):287-96.
- Opri E, Cernera S, Molina R, Eisinger RS, Cagle JN, Almeida L, et al. Chronic embedded cortico-thalamic closed-loop deep brain stimulation for the treatment of essential tremor. Science Translational Medicine. 2020;12(572):eaay7680.
- Rapinesi C, Kotzalidis GD, Ferracuti S, Sani G, Girardi P, Del Casale A. Brain Stimulation in Obsessive-Compulsive Disorder (OCD): A Systematic Review. Current Neuropharmacology. 2019;17(8):787-807.

- Deep Brain Stimulation for Treatment of Parkinson's Disease Deep brain stimulation, Parkinson's disease, subthalamic nucleus, stereotactic surgery. The Internet Journal of Neuromonitoring. 2012;7(1).
- Tsering D, Tochen L, Lavenstein B, Reddy SK, Granader Y, Keating RF, et al. Considerations in Deep Brain Stimulation (DBS) for pediatric secondary dystonia. Child's Nervous System. 2017;33(4):631-7.
- Holtzheimer PE, Husain MM, Lisanby SH, Taylor SF, Whitworth LA, McClintock S, et al. Subcallosal cingulate deep brain stimulation for treatment-resistant depression: A multisite, randomised, sham-controlled trial. The Lancet Psychiatry. 2017;4(11):839-49. Available from: https://www.ncbi.nlm.nih.gov/pubmed/289889 04
- 81. Jafari Z, Kolb BE, Mohajerani MH. Neural oscillations and brain stimulation in Alzheimer's disease. Progress in Neurobiology. 2020;194:101878.
- Jakobs M, Lee DJ, Lozano AM. Modifying the progression of Alzheimer's and Parkinson's disease with deep brain stimulation. Neuropharmacology. 2019;107860.
- Knotkova H, Hamani C, Sivanesan E, Beuffe MFEL, Moon JY, Cohen SP, et al. Neuromodulation for chronic pain. The Lancet. 2021;397(10289):2111-24. Available from: https://www.thelancet.com/pdfs/journals/lancet/PIIS0140-6736(21)00794-7. pdf
- Kundu B, Brock AA, Englot DJ, Butson CR, Rolston JD. Deep brain stimulation for the treatment of disorders of consciousness and cognition in traumatic brain injury patients: A review. Neurosurgical Focus. 2018;45(2):E14.
- Luigjes J, Segrave R, De Joode N, Figee M, Denys D. Efficacy of Invasive and Non-invasive Brain Modulation Interventions for Addiction. Neuropsychology Review. 2018;29(1):116-38.
- Wang TR, Moosa S, Dallapiazza RF, Elias WJ, Lynch WJ. Deep brain stimulation for the treatment of drug addiction. Neurosurgical Focus. 2018;45(2):E11. Available from:
- 87. https://thejns.org/focus/view/journals/neurosurg-focus/45/2/article-pE11.xml
- Zhou C, Zhang H, Qin Y, Tian T, Xu B, Chen J, et al. A systematic review and meta-analysis of deep brain stimulation in treatment-resistant depression. Progress in Neuro-Psychopharmacology and Biological Psychiatry. 2018;82:224-32. Available f rom: https://pubmed.ncbi.nlm.nih.gov/29146474/
- Rovira-Esteban L, Gunduz-Cinar O, Bukalo O, Limoges A, Brockway E, Müller K, et al. Excitation of Diverse Classes of Cholecystokinin Interneurons in the Basal Amygdala Facilitates Fear Extinction. Eneuro. 2019;6(6):ENEURO.0220-19.2019.
- Thankachan S, Katsuki F, McKenna JT, Yang C, Shukla C, Deisseroth K, et al. Thalamic Reticular Nucleus Parvalbumin Neurons Regulate Sleep Spindles and Electrophysiological Aspects of Schizophrenia in Mice. Scientific Reports. 2019;9(1):1-6.
- Spix TA, Nanivadekar S, Toong N, Kaplow IM, Isett BR, Goksen Y, et al. Population-specific neuromodulation prolongs therapeutic benefits of deep brain stimulation. Science. 2021;374(6564):201-6.
- 92. Gittis AH, Yttri EA. Translating Insights from Optogenetics to Therapies for Parkinson's Disease. Current Opinion in Biomedical Engineering. 2018;8:14-9. Available from: htt ps://www.ncbi.nlm.nih.gov/pmc/articles/PMC6941740/
- Lüscher C, Pascoli V, Creed M. Optogenetic dissection of neural circuitry: From synaptic causalities to blue prints for novel treatments of behavioral diseases. Current Opinion in Neurobiology. 2015;35:95-100.
- Lüscher C, Pollak P. Optogenetically inspired deep brain stimulation: Linking basic with clinical research. Swiss Medical Weekly. 2016;146(1314).
- 95. Vedam-Mai V, Deisseroth K, Giordano J, Lazaro-Munoz G, Chiong W, Suthana N, et al. Proceedings of the Eighth Annual Deep Brain Stimulation Think Tank: Advances in Optogenetics, Ethical Issues Affecting DBS Research, Neuromodulatory Approaches for Depression, Adaptive Neurostimulation, and Emerging DBS Technologies. Frontiers in Human Neuroscience. 2021;15:765150.
- Baker CK, Flannery JG. Innovative Optogenetic Strategies for Vision Restoration. Frontiers in Cellular Neuroscience. 2018;12:316.
- Bruchas MR, Roth BL. New Technologies for Elucidating Opioid Receptor Function. Trends in Pharmacological Sciences. 2016;37(4):279-89.
- Mukherjee A, Repina NA, Schaffer DV, Kane RS. Optogenetic tools for cell biological applications. Journal of Thoracic Disease. 2017;9(12):4867-70. Available from: https:/ /www.ncbi.nlm.nih.gov/pmc/articles/PMC5757028/
- Rost BR, Schneider-Warme F, Schmitz D, Hegemann P. Optogenetic Tools for Subcellular Applications in Neuroscience. Neuron. 2017;96(3):572-603. Available from: https://www.cell.com/neuron/pdf/S0896-6273(17)30916-9.pdf
- Shaaya M, Fauser J, Karginov AV. Optogenetics: The Art of Illuminating Complex Signaling Pathways. Physiology. 2021;36(1):52-60.
- 101. Iyer SM, Delp SL. Optogenetic Regeneration. Science. 2014;344(6179):44-5.
- Nihongaki Y, Kawano F, Nakajima T, Sato M. Photoactivatable CRISPR-Cas9 for optogenetic genome editing. Nature Biotechnology. 2015;33(7):755-60.
- Xu XM, Ordaz J, Wu W. Optogenetics and its application in neural degeneration and regeneration. Neural Regeneration Research. 2017;12(8):1197. Available from: https: //www.ncbi.nlm.nih.gov/pmc/articles/PMC5607808/

Cite this article: Figueiredo D, Sanghavi P. Bibliometric Analysis of Published Literature Utilizing Optogenetics as a Technique in Neuroscience: 2010–2020. J Scientometric Res. 2023;12(1)144-75.

| Ranking | Author name | Total document count | Year of 1st publication | Scopus Author ID | Total publication | <i>h</i> -index | Total citations |
|---------|----------------|----------------------------|-------------------------|---------------------|----------------------|-----------------|--------------------|
| 1 | Deisseroth, K. | 59 | 1990 | 6603640862 | 398 | 135 | 68179 |
| 2 | Boyden, E.S. | 26 | 2000 | 35291447700 | 208 | 67 | 19151 |
| 3 | Yamanaka, A. | 17 | 1997 | 55319368300 | 136 | 42 | 7084 |
| 4 | Kravitz, A.V. | 16 | 2004 | 6602759239 | 60 | 23 | 3403 |
| 5 | Lee, J.H. | 16 | 2002 | 57196135833 | 38 | 20 | 2194 |
| 6 | Murphy, T.H. | 15 | 1986 | 7401632487 | 175 | 59 | 12694 |
| 7 | Nestler, E.J. | 15 | 1977 | 7102006227 | 692 | 156 | 92273 |
| 8 | Stuber, G.D. | 15 | 2000 | 7007142142 | 105 | 48 | 10688 |
| 9 | BuzsÃiki, G. | 14 | 1973 | 7006676856 | 390 | 132 | 70008 |
| 10 | Kreitzer, A.C. | 14 | 2000 | 6507247826 | 62 | 45 | 10304 |

Table 1: Author details (Name and Scopus ID), optogenetic related document count, year of 1st publication, total publication count, h-index and total citations.

Table 2: Top cited article by the most productive authors in the field of optogenetics.

| Ranking | Author name | Most cited article in optogenetics | Times cited | Year | Journal |
|---------|----------------|--|-------------|------|--------------------------------|
| 1 | Deisseroth, K. | Optogenetics in Neural Systems | 1185 | 2011 | Neuron |
| 2 | Boyden, E.S. | A toolbox of Cre-dependent optogenetic transgenic mice for light-induced activation and silencing | 690 | 2012 | Nature Neuroscience |
| 3 | Yamanaka, A. | Acute optogenetic silencing of orexin/hypocretin neurons induces slow-wave sleep in mice | 166 | 2011 | Journal of Neuroscience |
| 4 | Kravitz, A.V. | Distinct roles for direct and indirect pathway striatal neurons in reinforcement | 533 | 2012 | Nature Neuroscience |
| 5 | Lee, J.H. | Frequency-selective control of cortical and subcortical networks by central thalamus | 66 | 2015 | eLife |
| 6 | Murphy, T.H. | Distinct Cortical Circuit Mechanisms for Complex Forelimb Movement and Motor Map Topography | 92 | 2012 | Neuron |
| 7 | Nestler, E.J. | The brain reward circuitry in mood disorders | 865 | 2013 | Nature Reviews Neuroscience |
| 8 | Stuber, G.D. | Optogenetic interrogation of dopaminergic modulation of the multiple phases of reward-seeking behavior | 232 | 2011 | Journal of Neuroscience |
| 9 | Buzsaki, G. | A toolbox of Cre-dependent optogenetic transgenic mice for light-induced activation and silencing | 690 | 2012 | Nature Neuroscience |
| 10 | Kreitzer, A.C. | Distinct roles for direct and indirect pathway striatal neurons in reinforcement | 533 | 2012 | Nature Neuroscience |

| Journal | Total Publications (TP) (2020) | Total Citations (TC) (2016-2019) | Cite Score 2019 | IF (Impact Factor) | <i>h</i> -index (2020) |
|------------------------------------|-----------------------------------|--|--------------------|-----------------------|---------------------------|
| Journal of Neuroscience | 3561 | 37553 | 10.5 | 5.6 | 14 |
| Neuron | 1448 | 36000 | 24.9 | 14.415 | 21 |
| Nature Neuroscience | 779 | 25093 | 32.2 | 20.0717 | 22 |
| Elife | 5574 | 60463 | 10.8 | 7.08 | 27 |
| Frontiers in Neural Circuits | 405 | 2373 | 5.9 | 3.156 | 5 |
| Eneuro | 1054 | 5211 | 4.9 | 3.544 | 7 |
| Journal of Neurophysiology | 1809 | 8132 | 4.5 | 2.225 | 6 |
| Frontiers in Cellular Neuroscience | 1705 | 9131 | 5.4 | 3.921 | 11 |
| Current Opinion in Neurobiology | 595 | 6422 | 10.8 | 6.528 | 11 |
| Journal of Visualized Experiments | 5213 | 9042 | 1.7 | 1.2 | 6 |

Table 3: Top 10 journals with respect to Total Publications, Total Citations, Cite Score 2019, IF (Impact Factor), and h-index (2020).

Table 4: Top 10 journals with respect to most cited articles and times-cited in 2020.

| Journal | Most cited article (2020) | Times cited (2020) |
|---------------------------------------|--|-----------------------|
| Journal of Neuroscience | Microglia actively remodel adult hippocampal neurogenesis through the phagocytosis secretome. | 51 |
| Neuron | TREM2 Regulates Microglial Cholesterol Metabolism upon Chronic Phagocytic Challenge. | 64 |
| Nature Neuroscience | Lipid-droplet-accumulating microglia represent a dysfunctional and proinflammatory state in the aging brain. | 85 |
| Elife | SARS-CoV-2 (COVID-19) by the numbers. | 149 |
| Frontiers in Neural Circuits | Multiplex Neural Circuit Tracing With G-Deleted Rabies Viral Vectors. | 10 |
| Eneuro | High-fat diet-induced obesity causes sex-specific deficits in adult hippocampal neurogenesis in mice. | 13 |
| Journal of Neurophysiology | Parallel distributed networks dissociate episodic and social functions within the individual. | 22 |
| Frontiers in Cellular Neuroscience | Cellular Senescence in Neurodegenerative Diseases. | 30 |
| Current Opinion in Neurobiology | Deep learning tools for the measurement of animal behavior in neuroscience. | 31 |
| Journal of Visualized Experiments | Measurement of the hepatic venous pressure gradient and transjugular liver biopsy. | 10 |